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(54) **YEAST STRAIN AND METHOD FOR USING THE SAME TO PRODUCE NICOTINAMIDE RIBOSIDE**

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**A23L 1/28** (2006.01)  
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(52) **U.S. Cl.** .... 435/61; 435/256.1; 435/71.1; 435/254.1

(58) **Field of Classification Search** ..... None  
See application file for complete search history.

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(57) **ABSTRACT**

The present invention embraces a fungal strain deficient in nicotinamide riboside import and salvage and use thereof for producing nicotinamide riboside. Methods for producing nicotinamide riboside and a nicotinamide riboside-supplemented food product using the strain of the invention are also provided.

**11 Claims, 1 Drawing Sheet**

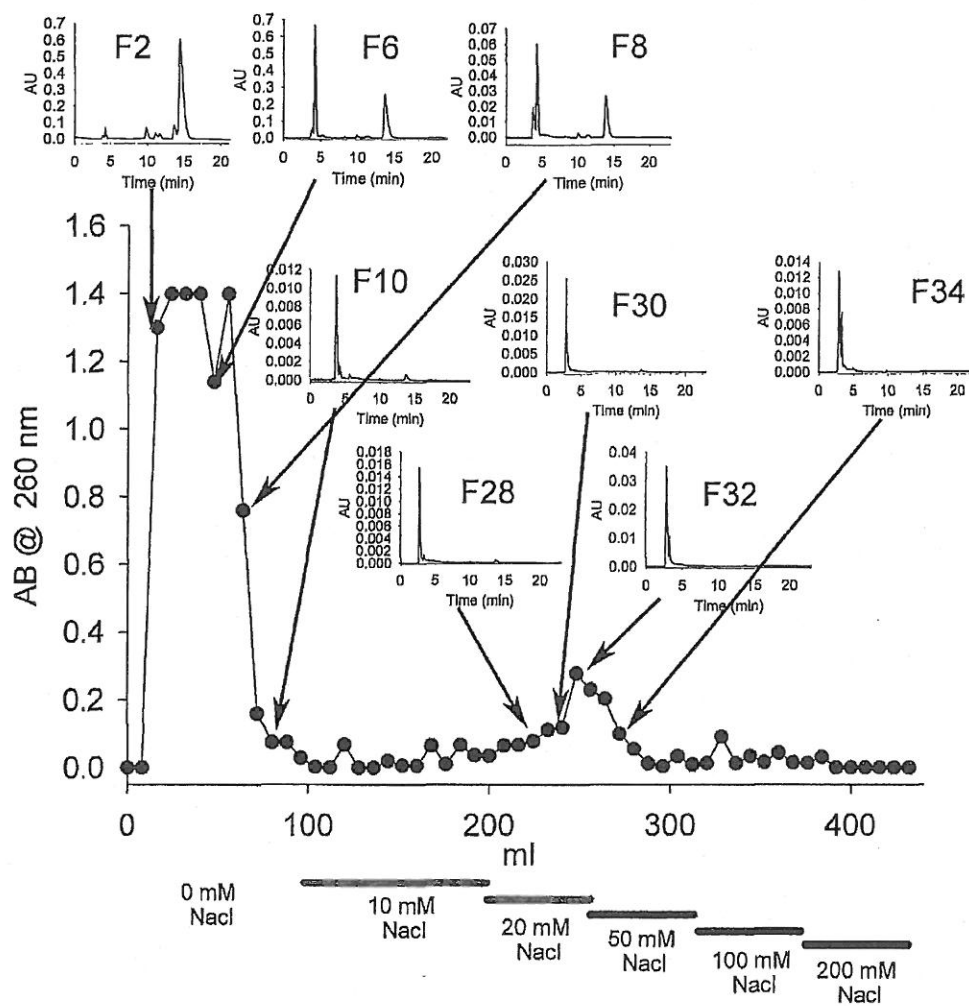


FIG. 1

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# YEAST STRAIN AND METHOD FOR USING THE SAME TO PRODUCE NICOTINAMIDE RIBOSIDE

This application is a continuation-in-part of U.S. patent application Ser. No. 11/542,832, filed Oct. 4, 2006, which is a divisional of U.S. patent application Ser. No. 11/113,701, filed Apr. 25, 2005 now abandoned, which is a continuation-in-part of PCT/US2005/004337, filed Feb. 9, 2005, which claims benefit of U.S. Provisional Patent Application Ser. No. 60/543,347, filed Feb. 10, 2004, the contents of which are incorporated herein by reference in their entireties.

This invention was made in the course of research sponsored by the National Science Foundation, grant number MCB-0822581, and the National Institutes of Health, grant number T32GM008704. The U.S. government has certain rights in this invention.

## INTRODUCTION

### Background of the Invention

Nicotinic acid (NA), nicotinamide (Nam) and nicotinamide riboside (NR) constitute three salvageable NAD<sup>+</sup> precursor vitamins in yeast. NA is imported by the high affinity major facilitator superfamily (MSF) type transporter Tna1 (Llorente & Dujon (2000) *FEBS Lett.* 475:237-41; Klebl, et al. (2000) *FEBS Lett.* 481:86-7). However, not all NA import is Tna1-dependent and at concentrations above 1  $\mu$ M NA, Tna1-independent import is detectable (Llorente & Dujon (2000) *supra*). NA is converted to NAD<sup>+</sup> via the 3-step Preiss-Handler pathway (Preiss & Handler (1958) *J. Biol. Chem.* 233:488-92; Preiss & Handler (1958) *J. Biol. Chem.* 233:493-500). Nam is converted to NA by the nicotinamidase (Pnc1) (Ghislain, et al. (2002) *Yeast* 19:215-24; Anderson, et al. (2003) *Nature* 423:181-5), for entry into Preiss-Handler salvage. A Nam transporter has not been identified.

## SUMMARY OF THE INVENTION

The present invention features an isolated fungal strain deficient in nicotinamide riboside import and salvage. In one embodiment, the strain does not express Nicotinamide Riboside Kinase 1 (NrK1), Uridine Hydrolase 1 (Urh1), Purine Nucleoside Phosphorylase (Pnp1), and Nicotinamide Riboside Transporter 1 (Nrt1). In another embodiment, the strain secretes at least 8 mg/L nicotinamide riboside. In a further embodiment, the fungus is selected from the group consisting of *Saccharomyces*, *Schizosaccharomyces*, *Kluveromyces*, *Aspergillus* and *Pichia*. In a specific embodiment, the fungus is *Saccharomyces cerevisiae*.

The present invention also embraces a method for producing nicotinamide riboside by culturing the fungal strain of the invention in culture medium and recovering nicotinamide riboside from the medium. In one embodiment, the culture medium further includes nicotinic acid or nicotinamide. In another embodiment, the fungal strain is cultured to an optical density of at least 3. In a particular embodiment, the nicotinamide riboside is recovered by solubilizing nicotinamide riboside from the medium with methanol and subjecting the nicotinamide riboside to column chromatography.

A method for producing a nicotinamide riboside-supplemented food product is also provided. According to this method, a fermentable substrate is fermented in the presence of the fungal strain of the invention thereby producing a nicotinamide riboside supplemented food product. A nicotinamide riboside supplemented food product fermented in the

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presence of the fungal strain of the invention is also provided. In some embodiments, the food product is wine, beer, cider, kvass, root beer, soy sauce or bread.

## BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows the purification of NR from PAB076-conditioned media. Media collected from PAB076 grown to optical density at 600 nm ( $OD_{600nm}$ ) of 60 in 2 $\times$ YPD and supplemented with 5 mM NA was cleaned and concentrated by lyophilization followed by resuspension in cold methanol. This material was then loaded directly onto the SP-SEPHADEX resin. FIG. 1 shows the measured absorbance of fractions collected from preparative SP-SEPHADEX chromatography. Salt concentration is depicted below the x-axis. An HPLC chromatogram of each fraction was obtained and selected traces are included as the eight smallest inlays. NR eluted between fraction 27 and 36.

## DETAILED DESCRIPTION OF THE INVENTION

NR is converted into NAD<sup>+</sup> through two distinct pathways. The first pathway utilizes the NR kinase, NrK1, to produce nicotinamide mononucleotide, which is then converted into NAD<sup>+</sup>. The second pathway cleaves NR into Nam and a ribose, by exploiting two independently acting enzymes uridine hydrolase 1 (Urh1) and purine nucleoside phosphorylase (Pnp1). Jointly these pathways are described as the NR salvage pathways and they feed into the NAD<sup>+</sup> cycle in two places.

It has now been shown that mutants which are deficient in NR salvage (i.e., nrk1 urh1 pnp1) can export NR in an Nrt-independent manner and support the growth of the NR auxotroph, qns1. More significantly, deletion of Nrt1 in a nrk1 urh1 pnp1 strain actually leads to increased extracellular NR accumulation. Moreover, NA or nicotinamide supplementation of a nrk1 urh1 pnp1 nrt1 strain increases NR yield from the strain. Accordingly, the present invention embraces a fungal strain deficient in the salvage and import of NR and use of said strain as a source for the production of NR. In addition, the invention provides a simple and scalable extraction method for inexpensively obtaining NR. Fungal strains of the present invention find application in large-scale production of NR as well as in the processes for fermenting of bread, soy, wine, beer, cider, kvass, root beer and other beverages, thereby providing added value of high nicotinamide riboside content. The nicotinamide riboside produced and isolated according to the present invention finds use in dietary supplement and pharmaceutical compositions for the prevention and treatment of a disease or condition associated with the nicotinamide riboside kinase pathway of NAD<sup>+</sup> biosynthesis.

As indicated, the present invention embraces an isolated fungal strain deficient in nicotinamide riboside import and salvage. For the purposes of the present invention, a "fungal strain deficient in nicotinamide riboside import and salvage" is a strain that fails to import nicotinamide into the cytoplasm and also fails to utilize nicotinamide riboside as a NAD<sup>+</sup> precursor. In one embodiment, the fungal strain is produced by destroying or deleting by knocking out one or more genes involved in import and salvage of NR. Such gene deletions or disruptions are routinely practiced in the art and any conventional method, including those exemplified herein, can be employed.

In accordance with particular embodiments, the fungal strain of the invention does not express Nicotinamide Riboside Kinase 1 (NrK1), Uridine Hydrolase 1 (Urh1), Purine Nucleoside Phosphorylase 1 (Pnp1), and Nicotinamide Ribo-

side Transporter 1 (Nrt1). Genes encoding these proteins are known in the art and available from databases such as NCBI Entrez Nucleotide database, the *Saccharomyces* Genome Database, and the *Schizosaccharomyces pombe* genome project. For example, Nrk1 is provided under GENBANK accession nos. NP\_014270 (SEQ ID NO:13, *S. cerevisiae*), NP\_595603 (SEQ ID NO:14, *S. pombe*), XP\_456163 (SEQ ID NO:15, *Kluveromyces lactis*), XP\_001820220 (SEQ ID NO:16, *Aspergillus oryzae*), and XP\_001386700 (SEQ ID NO:17, *Pichia stipitis*). Similarly, Urh1 is provided under GENBANK accession nos. NP\_010688 (SEQ ID NO:18, *S. cerevisiae*), NP\_593725 (SEQ ID NO:19, *S. pombe*), XP\_452497 (SEQ ID NO:20, *K. lactis*), XP\_001816861 (SEQ ID NO:21, *A. oryzae*), and XP\_001384876 (SEQ ID NO:22, *P. stipitis*). Pnp1 is provided under GENBANK accession nos. NP\_013310 (SEQ ID NO:23, *S. cerevisiae*), NP\_593927 (SEQ ID NO:24, *S. pombe*), and XP\_452943 (SEQ ID NO:25, *K. lactis*). In addition, Nrt1 is provided under GENBANK Accession Nos. NP\_014714 (SEQ ID NO:26, *S. cerevisiae*), NP\_595061 (SEQ ID NO:27, *S. pombe*), XP\_453096 (SEQ ID NO:28, *K. lactis*), XP\_001821563 (SEQ ID NO:29, *A. oryzae*), and XP\_001383412 (SEQ ID NO:30, *P. stipitis*). Using these known sequences, the skilled artisan can readily disrupt or knockout the genes of interest to obtain a fungal strain deficient in NR transport and salvage. Strains with the desired gene knockouts or deletions can be identified by routine screens including, but not limited to, Southern blot analysis, RT-PCR, northern blot analysis, western blot analysis and the like.

In certain embodiments, the fungal strain of the present invention is used in the production of pharmaceuticals or in food fermentation, e.g., in the production of bread, wine, beer, cider, kvass, root beer, cheese, or soy sauce. In accordance with such embodiments, the fungal strain of the invention is selected from the genus *Saccharomyces*, *Schizosaccharomyces*, *Kluveromyces*, *Pichia*, or *Aspergillus* (e.g., *A. oryzae* or *A. sojae*). In particular embodiments, the fungal strain is a yeast, e.g., a fungus of the genus *Saccharomyces* (e.g., *S. cerevisiae*, *S. bayanus*, *S. boulardii*, *S. pastorianus*, *S. rouxii* and *S. uvarum*), *Schizosaccharomyces* (e.g., *S. pombe*), *Kluveromyces* (e.g., *K. lactis* and *K. fragilis*) and *Pichia*. In particular embodiments, the fungus is *Saccharomyces cerevisiae*.

Unexpectedly, by blocking NR uptake and salvage, the strain of this invention secretes at least 4.0  $\mu\text{M}$  or 8 mg/L of nicotinamide riboside into the culture medium; a 40-fold increase over production of nicotinamide riboside in a wild-type strain. Furthermore, supplementation of the culture medium with either nicotinic acid or nicotinamide increases nicotinamide riboside production to as much as 7-8  $\mu\text{M}$ , wherein even higher amounts of nicotinamide riboside are produced when the cells are cultured to extremely high densities. For example, *S. cerevisiae* grown to an  $\text{OD}_{600\text{nm}}$  of 60 in 2xYPD+5 mM NA was capable of producing 28  $\mu\text{M}$  nicotinamide riboside.

Thus, given the significant amount of nicotinamide riboside secreted by a fungal strain deficient in NR transport and salvage, the present also features a method for producing nicotinamide riboside by culturing the fungal strain of the invention in growth medium and recovering the methanol-solubilized nicotinamide riboside from the medium. In accordance with this method, the fungal strain is cultured in a fermentation, culture, or growth medium for production of nicotinamide riboside. An appropriate, or effective, culture medium refers to any medium in which a fungal strain of the present invention, when cultured, is capable of producing

nicotinamide riboside. Such a medium is typically an aqueous medium composed of assimilable carbon, nitrogen and phosphate sources. Such a medium can also include appropriate salts, minerals, metals, and other nutrients. It should be recognized, however, that a variety of fermentation conditions are suitable and can be selected by those skilled in the art based upon art recognized culture conditions and the teachings of the present disclosure. In this regard, particular embodiments embrace the addition of nicotinamide or nicotinic acid to the culture medium. In other embodiments, the culture medium is formulated to support extremely high densities of cells, i.e., an  $\text{OD}_{600\text{nm}}$  of at least 3.

Depending on the result to be achieved, the fungus can be cultured under anaerobic (deficient in oxygen) as well as aerobic (oxygenated) conditions. Under aerobic conditions, microorganisms such as yeast cells can break down sugars to end products such as  $\text{CO}_2$  and  $\text{H}_2\text{O}$ . Under anaerobic conditions, yeast cells utilize an alternative pathway to produce  $\text{CO}_2$  and ethanol. The fermentation reaction of the present invention is preferably anaerobic, i.e., partially or completely deficient in oxygen. Fermentation can also be used to refer to the bulk growth of microorganisms on a growth medium where no distinction is made between aerobic and anaerobic metabolism.

Fungal strains of the present invention can be cultured in conventional fermentation modes, which include, but are not limited to, batch, fed-batch, cell recycle, and continuous. In a fed-batch mode, when during fermentation some of the components of the medium are depleted, it may be possible to initiate the fermentation with relatively high concentrations of such components so that growth is supported for a period of time before additions are required. The preferred ranges of these components are maintained throughout the fermentation by making additions as levels are depleted by fermentation. Levels of components in the fermentation medium can be monitored by, for example, sampling the fermentation medium periodically and assaying for concentrations. Alternatively, once a standard fermentation procedure is developed, additions can be made at timed intervals corresponding to known levels at particular times throughout the fermentation. The additions to the fermentor may be made under the control of a computer in response to fermentor conditions or by a preprogrammed schedule. Moreover, to avoid introduction of foreign microorganisms into the fermentation medium, addition is performed using aseptic addition methods, as are known in the art. In addition, a small amount of anti-foaming agent may be added during the fermentation, or anti-foaming device may be employed.

In particular embodiments, recovery of the nicotinamide riboside from the culture medium is achieved by a simple, inexpensive process. The process involves solubilizing the nicotinamide riboside from the medium with methanol leaving behind a methanol-insoluble pellet; and subjecting the nicotinamide riboside to column chromatography to isolate the nicotinamide riboside from other contaminants. To facilitate the solubilization step, the culture medium can be concentrated, e.g., by lyophilization (freeze-drying) or roto-evaporation. In addition to the SP-SEPHADEX column chromatography exemplified herein, nicotinamide riboside can alternatively or also be purified by solid phase extraction, porous graphitic carbon or hydrophilic interaction chromatography. It is contemplated that the number and types of chromatographic columns employed will be dependent on the final use of the nicotinamide riboside and the level of purification desired.

In so far as yeast and other fungi are routinely used in the production of food products, the present invention also



embraces a method for producing a nicotinamide riboside supplemented food product by providing a fermentable substrate and fermenting the fermentable substrate in the presence of the fungal strain of the invention. Food products, which can be produced in accordance with the method of this invention include, but are not limited to, bread, cheese, wine, beer, cider, kvass, root beer, or other beverages. As such, a fermentable substrate is intended to include any substratum which, when fermented, produces the above-referenced food products. Fermentable substrates include, but are not limited to, vegetables, oat, wheat, barley, millet, rice, rye, sorghum, potato, fruits, fruit juices, and the like.

Nicotinic acid is an effective agent in controlling low-density lipoprotein cholesterol, increasing high-density lipoprotein cholesterol, and reducing triglyceride and lipoprotein (a) levels in humans (see, e.g., Miller (2003) *Mayo Clin. Proc.* 78(6):735-42). Though nicotinic acid treatment effects all of the key lipids in the desirable direction and has been shown to reduce mortality in target populations, its use is limited because of a side effect of heat and redness termed flushing, which is significantly effected by the nature of formulation. Further, nicotinamide protects against stroke injury in model systems, due to multiple mechanisms including increasing mitochondrial NAD<sup>+</sup> levels and inhibiting PARP (Klaidman, et al. (2003) *Pharmacology* 69(3):150-7). Altered levels of NAD<sup>+</sup> precursors have been shown to effect the regulation of a number of genes and lifespan in yeast (Anderson, et al. (2003) *Nature* 423(6936):181-5).

NAD<sup>+</sup> administration and NMN adenyltransferase (Nmnat1) expression have also been shown to protect neurons from axonal degeneration (Araki, et al. (2004) *Science* 305:1010-1013). Because nicotinamide riboside is a soluble, transportable nucleoside precursor of NAD<sup>+</sup>, nicotinamide riboside can be used to protect against axonopathies such as those that occur in Alzheimer's Disease, Parkinson's Disease and Multiple Sclerosis. As such administration of nicotinamide riboside or a nicotinamide riboside supplemented-food product could also protect against axonal degeneration.

NMN adenyltransferase overexpression has been shown to protect neurons from the axonopathies that develop with ischemia and toxin exposure, including vincristine treatment (Araki, et al. (2004) *Science* 305:1010-1013). Vincristine is one of many chemotherapeutic agents whose use is limited by neurotoxicity. Thus, administration of nicotinamide riboside or a nicotinamide riboside supplemented-food product could be used to protect against neurotoxicity before, during or after cytotoxic chemotherapy.

Further, conversion of benign *Candida glabrata* to the adhesive, infective form is dependent upon the expression of EPA genes encoding adhesins whose expression is mediated by NAD<sup>+</sup> limitation, which leads to defective Sir2-dependent silencing of these genes (Domergue, et al. (March 2005) *Science*, 10.1126/science.1108640). Treatment with nicotinic acid reduces expression of adhesins and increasing nicotinic acid in mouse chow reduces urinary tract infection by *Candida glabrata*. Thus, nicotinamide riboside or a nicotinamide riboside-supplemented food product can be used in the treatment of fungal infections, in particular, those of *Candida* species by preventing expression of adhesins.

Accordingly, the nicotinamide riboside or a nicotinamide riboside-supplemented food product of this invention could have therapeutic value in improving plasma lipid profiles, preventing stroke, providing neuroprotection with chemotherapy treatment, treating fungal infections, preventing or reducing neurodegeneration, or in prolonging health and well-being. Thus, the present invention is further a method for preventing or treating a disease or condition associated with

the nicotinamide riboside kinase pathway of NAD<sup>+</sup>-biosynthesis by administering an effective amount of a nicotinamide riboside composition. Diseases or conditions which typically have altered levels of NAD<sup>+</sup> or NAD<sup>+</sup> precursors or could benefit from increased NAD<sup>+</sup>-biosynthesis by treatment with nicotinamide riboside include, but are not limited to, lipid disorders (e.g., dyslipidemia, hypercholesterolaemia or hyperlipidemia), stroke, neurodegenerative diseases (e.g., Alzheimer's, Parkinsons and Multiple Sclerosis), neurotoxicity as observed with chemotherapies, *Candida glabrata* infection, and the general health declines associated with aging. Such diseases and conditions can be prevented or treated by diet supplementation or providing a therapeutic treatment regime with a nicotinamide riboside composition.

An effective amount of nicotinamide riboside is one which prevents, reduces, alleviates or eliminates the signs or symptoms of the disease or condition being prevented or treated and will vary with the disease or condition. Such signs or symptoms can be evaluated by the skilled clinician before and after treatment with the nicotinamide riboside to evaluate the effectiveness of the treatment regime and dosages can be adjusted accordingly.

The nicotinamide riboside produced in accordance with the method of the invention can be conveniently used or administered in a composition containing the active agent in combination with a pharmaceutically acceptable carrier. Such compositions can be prepared by methods and contain carriers which are well-known in the art. A generally recognized compendium of such methods and ingredients is Remington: The Science and Practice of Pharmacy, Alfonso R. Gennaro, editor, 20th ed. Lippincott Williams & Wilkins: Philadelphia, Pa., 2000. A carrier, pharmaceutically acceptable carrier, or vehicle, such as a liquid or solid filler, diluent, excipient, or solvent encapsulating material, is involved in carrying or transporting the subject compound from one organ, or portion of the body, to another organ, or portion of the body. Each carrier must be acceptable in the sense of being compatible with the other ingredients of the formulation and not injurious to the patient.

Examples of materials which can serve as carriers include sugars, such as lactose, glucose and sucrose; starches, such as corn starch and potato starch; cellulose, and its derivatives, such as sodium carboxymethyl cellulose, ethyl cellulose and cellulose acetate; powdered tragacanth; malt; gelatin; talc; excipients, such as cocoa butter and suppository waxes; oils, such as peanut oil, cottonseed oil, safflower oil, sesame oil, olive oil, corn oil and soybean oil; glycols, such as propylene glycol; polyols, such as glycerin, sorbitol, mannitol and polyethylene glycol; esters, such as ethyl oleate and ethyl laurate; agar; buffering agents, such as magnesium hydroxide and aluminum hydroxide; alginic acid; pyrogen-free water; isotonic saline; Ringer's solution; ethyl alcohol; pH buffered solutions; polyesters, polycarbonates and/or polyanhydrides; and other non-toxic compatible substances employed in formulations. Wetting agents, emulsifiers and lubricants, such as sodium lauryl sulfate and magnesium stearate, as well as coloring agents, release agents, coating agents, sweetening, flavoring and perfuming agents, preservatives and antioxidants can also be present in the compositions.

Nicotinamide riboside produced in accordance with the method of the invention can be administered via any route include, but not limited to, oral, rectal, topical, buccal (e.g., sub-lingual), vaginal, parenteral (e.g., subcutaneous, intramuscular including skeletal muscle, cardiac muscle, diaphragm muscle and smooth muscle, intradermal, intravenous, intraperitoneal), topical (i.e., both skin and mucosal surfaces, including airway surfaces), intranasal, transdermal, intraar-

ticular, intrathecal and inhalation administration, administration to the liver by intraportal delivery, as well as direct organ injection (e.g., into the liver, into the brain for delivery to the central nervous system). The most suitable route in any given case will depend on the nature and severity of the condition being treated and on the nature of the particular compound which is being used.

A physician or veterinarian having ordinary skill in the art can readily determine and prescribe the effective amount of the pharmaceutical composition required for prevention or treatment in an animal subject such as a human, agriculturally-important animal, pet or zoological animal.

In addition to the specific fungal strains disclosed herein, it is expected that these fungal strains may be further manipulated to achieve other desirable characteristics, or even higher specific yields of fermentation products. For example, selection of strains by passaging the strains of the present invention on medium containing a particular substrate of interest may result in improved fungi with enhanced fermentation rates.

The invention is described in greater detail by the following non-limiting examples.

### Example 1

#### Materials and Methods

**Yeast Strains and Medium.** All *Saccharomyces cerevisiae* strains used in this study were derivatives of the common wild-type strain, BY4742. Construction of single deletion strains was according to established methods (Winzeler, et al. (1999) *Science* 285:901-6). Additional deletions were created by direct transformation with PCR products (Brachmann, et al. (1998) *Yeast* 14:115-32). Primers employed in the PCR reactions are listed in Table 1.

TABLE 1

Primer	Sequence (5' to 3')	SEQ ID NO:
14050	gctctagacagacaagtggtatgcataatcc	1
14051	cgggggtaccgatgtgctgtgactggg	2
14060	gccgctcgagcttcccgctatgtaataaagag	3
14061	cgccgatccgcatactctgtcaatttccttg	4
14121 NRT1 Deletion F	GAATTTATATTATTCTTTATTGTACTGATATCCCCATTATACTATCAAAAAAGGAC TTCAGCACCTGTGCGGTATTTACACCG	5
14122 NRT1 Deletion R	CTGTACAGATTTCAAATGAAGCGTTGAA GTTTCCTCTTTGTATATTTGAGATCTTCA TTTTATCAGATTGTACTGAGAGTGCA	6
14124 NRT1 Diagnostic F	CTAGTGTGTGCTACCGCTATTTGTTCTTCG	7
14124 NRT1 Diagnostic R	GCAGTCGAGGATCGATCTGGTAGTATTC	8
4750	AATAGCGTGC AAAAGCTATCGAAGTGTGA GCTAGAGTAGAACCTCAAAATAGATTGTA CTGAGAGTGCA	9
4751	CTAATCCTTACAAAGCTTGAATCTCTT GGCACACCCAGCTTAAAGGTCGTGCGGT ATTTCACACCG	10

TABLE 1-continued

Primer	Sequence (5' to 3')	SEQ ID NO:
14113	CTCTCCGAGCTCGGATTCTTTGTTCATCAGA CAACTTGTGTGAGTGG	11
14112	GTGCCCAAGCTTGTGTGCCAATGTAGCGTG GTTGCATG	12

pPAB01 was constructed by amplifying the PNP1 gene from wild-type yeast genomic DNA with primers 14061 and 14060. The PCR product was inserted into pRS416 with XhoI and BamHI. pPAB02 was constructed by amplifying the URH1 gene using primers 14051 and 14050. The PCR product was inserted into pRS416 with KpnI and XbaI. Plasmids were confirmed by DNA sequencing and used for construction of deletion strains.

A yeast strain carrying disruption of the NRK1 locus was made by transformation of the strain BY165-1d with the HIS3 marker introduced into disruption cassette by PCR with primers 4750 and 4751.

Plasmid pNRT1, carrying NRT1 under the control of its own promoter, was created by amplifying the gene from BY4742 DNA using primers 14112 and 14113. After digestion with SacI and HindIII, the product was inserted into pRS317.

Strains generated and used herein are listed in Table 2.

TABLE 2

Name	Genotype
B4742 <sup>a</sup>	MAT $\alpha$ his3 $\Delta$ 1 leu2 $\Delta$ 0 lys2 $\Delta$ 0 ura3 $\Delta$ 0
PAB011	BY4742 nrt1 $\Delta$ ::kanMX4
PAB038	BY4742 pnp1 $\Delta$ ::kanMX4 urh1::NAT nrk1 $\Delta$ ::HIS3
PAB075	BY4742 nrt1 $\Delta$ ::kanMX4 fun26 $\Delta$ ::URA3
PAB076	BY4742 pnp1 $\Delta$ ::kanMX4 urh1::NAT nrk1 $\Delta$ ::HIS3 nrt1 $\Delta$ ::URA3
PY165-d	qns1::URA3 pB175

<sup>a</sup>Brachmann, et al. (1998) *Yeast* 14: 115-32.

NA-free synthetic dextrose complete media (SDC) and its vitamin supplemented forms are described in the art (Wickham (1946) *J. Bacteriol.* 52:293-301). 2 $\times$ SDC and 2 $\times$ YPD were prepared as the more concentrated forms of the conventional preparation.

qns1 Bioassay. Strain BY165-1d, the chromosomal deletion of qns1 carrying the QNS1 plasmid pB175 (Bieganski, et al. (2003) *J. Biol. Chem.* 278:33049-33055), was plated on 5-FOA plates supplemented with NR to remove pB175. The resulting strain was cultured on NR containing media at all times. Conditioned media was prepared by incubating the specified yeast strain in the appropriate media. After 18 hours the cells were removed by centrifugation followed by filtration. The conditioned media was retained and mixed in a 1 to 1 ratio with fresh 2 $\times$ SDC. BY165-1d with no pB175 was incubated in the resulting media and growth was measured spectroscopically.

MALDI-MS NR Quantification. NR content in conditioned media was measured using MALDI-MS. Prior to measurement, [<sup>18</sup>O] NR was added to the media to a final concentration of 10  $\mu$ M as an internal standard. One microliter of the [<sup>18</sup>O] spiked samples was mixed with 1  $\mu$ l 2,5-Dihydroxy benzoic acid (DHB) matrix, and the mixture was allowed to air dry. The DHB matrix was composed of 50% acetonitrile saturated with DHB. MS spectra were collected on the ABI Voyager-DE Pro MALDI-TOF mass spectrometer and the ratio of the labeled standard to the unlabeled NR was used to determine the NR concentration.

HPLC Measurements. NA, Nam and NR were also measured using HPLC. Media samples were injected directly onto a Princeton SPHER-60 SAX 60A u (250×4.6 mm) column and separated by an isocratic run of 20 mM  $\text{KH}_2\text{PO}_4$ . Metabolites were detected spectroscopically at 260 nm and quantified by comparison to a standard curve.

NR Extraction. NR was extracted from 2×YPD. PAB076 was incubated in 500 ml of 2×YPD to an  $\text{OD}_{600\text{nm}}$  of 60 (~60 hours). The media was divided into 150 ml portions and frozen at  $-80^\circ\text{C}$ . As the first step in the purification process, the samples were lyophilized and resuspended in 25 ml of cold methanol. Cold methanol solubilized the NR but left the majority of the contaminants as a pellet after centrifugation. The methanol samples were then lyophilized again and resuspended in 5 ml of water. The aqueous samples were then run over a 10 ml SP-SEPHADEX column, and eluted using a stepped NaCl gradient. NR eluted at 25–50 mM NaCl. Fractions were analyzed using HPLC, and NR was confirmed using MALDI-MS and a biological  $\text{NAD}^+$  assay.

Biological  $\text{NAD}^+$  Assay. Yeast cultures were grown with agitation in 0.5 L cultures. During growth, the  $\text{OD}_{600\text{nm}}$  of 1:10 diluted cells were recorded and 20 ml cultural volumes were pelleted, washed with water, repelleted, and frozen at  $-80^\circ\text{C}$ . Cell pellets were extracted in 250 ml of ice-cold 1 M formic acid saturated with butanol. After 30 minutes, 62.5 ml of 100% (w/v) trichloroacetic acid was added to each extract, and the samples were allowed to precipitate on ice for 15 minutes. Samples were microcentrifuged for 5 minutes, and the acid soluble supernatants were recovered. Pellets were washed with 125 ml of 20% TCA and repelleted. First and second supernatants were pooled and measured volumetrically. In three 1 ml cuvettes, reactions were assembled containing 10 ml 5 mg/ml alcohol dehydrogenase (two samples) or 10 ml water (control sample), and this was followed by addition of 840 ml 360 mM Tris (pH 9.7), 240 mM lysine, 0.24% (v/v) EtOH, and 150 ml extract. After a 5 minute incubation at room temperature, the spectrophotometer was zeroed against the control sample for determining the alcohol dehydrogenase-dependent increase in absorbance at 340 nm of the duplicate reactions. Mean net absorbances were converted to molar  $\text{NAD}^+$  with the extinction coefficient of NADH ( $6220\text{M}^{-1}\text{cm}^{-1}$ ). Molar  $\text{NAD}^+$  in the cuvette was converted to molar  $\text{NAD}^+$  in the extract by a factor of 6.67. Moles of  $\text{NAD}^+$  in the extract were determined from the fraction of the extract assayed. To determine the intracellular volumes corresponding to the extracts and the corresponding intracellular  $\text{NAD}^+$  concentrations, a nonlinear conversion between the 1:10 diluted  $\text{OD}_{600\text{nm}}$  values and the cell number was used (Burke, et al. (2000) *Methods in Yeast Genetics*, Cold Spring Harbor, N.Y.: Cold Spring Harbor Press) and took the volume of a haploid cell to be  $7\times 10^{-14}$  (Sherman (1991) *Methods Enzymol.* 194:3–21). For cells grown in media containing nicotinic acid,  $\text{NAD}^+$  concentrations were determined, in duplicate, 6 to 18 times during the growth of a liquid culture. For cells grown in media without nicotinic acid, the cells were taken with 1:10 diluted  $\text{OD}_{600\text{nm}}$  values of 0.095–0.105, and the  $\text{NAD}^+$  concentrations were determined, in duplicate, from three to eight independent cultures.

#### Example 2

##### NR Export is Nrt1-Independent

In yeast, NR has activity as a *qns1*-bypassing and lifespan extending vitamin. It has also been found that NR is an intracellular and extracellular metabolite. On the basis of the discovery of the specific NR transporter, Nrt1 (YOR071C gene), it was of interest to determine whether this importer is responsible for the observed NR export activity.

The NR-non-salvaging genotype *nrt1 urh1 pnp1* (strain PAB038) exhibits reduced  $\text{NAD}^+$  levels and exports NR. To test whether Nrt1 is required for the export of NR, NRT1 was deleted in the PAB038 strain through homologous recombination using the URA3 marker to replace NRT1.

Extracellular NR is detectable using a *qns1* bioassay that relies on the NR auxotrophy of the *qns1* strain. In this assay, the strains being tested for NR export are grown overnight in SDC medium, at which point the cells are removed and the conditioned media is retained. The *qns1* strain is then incubated in medium containing equal measures of conditioned media and fresh 2×SDC. In this assay, the extent of *qns1* growth is proportional to the extracellular concentration of NR. Based on *qns1* growth, the *nrt1* deletion does not reduce extracellular NR. On the contrary NR levels are actually elevated. By comparison to SDC supplemented with purified NR, it was estimated that the NR-non-salvaging strain, PAB038, produced 1  $\mu\text{M}$  extracellular NR when incubated to an OD of 3, whereas the NR-non-salvaging and NR-non-importing strain, *nrt1 urh1 pnp1 nrt1* (PAB076), produced 2  $\mu\text{M}$  extracellular NR under the same growth conditions. The excess of extracellular NR in the *nrt1* mutant was apparently due to the fact that NR export was Nrt1-independent. The results of this analysis indicated that in strain PAB076, NR can be exported but not reabsorbed, resulting in higher accumulation of extracellular NR by the PAB076 strain.

#### Example 3

##### Increases in NR Yield

NR has potential to become an important vitamin for daily dietary supplementation and at higher levels a drug for the treatment of disorders like dyslipidemia. One of the hurdles to the development of NR as a product for human consumption has been the difficulty and expense of enzymatic or chemical synthesis. Nicotinamide riboside is costly to produce, largely because of the cost of blocked (i.e., acetylated or benzoylated) ribose used in its organic synthesis (Tanimori, et al. (2002) *Bioorg. Med. Chem.* 12:1135–1137). As such, improved NR export from yeast may provide a clean and simple biological alternative to the current modes of NR production. It was contemplated that one possible way to upregulate NR export would be to supplement yeast with the inexpensive  $\text{NAD}^+$  precursors NA or Nam. Niacin supplementation would have two potentially beneficial effects: first it would help replenish  $\text{NAD}^+$  lost in the synthesis of NR and second it would lead to the over expression of NR producing 5' nucleotidases.

Assaying the content of NR in media conditioned by PAB076 in the presence of 1 mM NA or Nam revealed that supplementation substantially increased the amount of NR produced as assayed by *qns1* growth. The extent of *qns1* growth was higher than the growth provided by 3  $\mu\text{M}$  NR, indicating that the concentration of NR in the conditioned media was at least 6  $\mu\text{M}$ .

The *qns1* bioassay is an effective method of detecting the presence of low amounts of NR in conditioned media but becomes nonlinear at high concentrations. To more accurately measure the extracellular concentration of NR, MALDI-MS was employed with an internal standard of [ $^{18}\text{O}$ ] NR at a concentration of 10  $\mu\text{M}$ . The concentration of NR in the media was determined from the ratio of the labeled standard to the unlabeled NR.

Using MS quantification, it was found that wild-type yeast had  $0.120\pm 0.4$   $\mu\text{M}$  NR, PAB038 (*pnp1 urh1 nrt1*) had  $1.2\pm 0.4$   $\mu\text{M}$  NR and PAB076 (*pnp1 urh1 nrt1*) had  $4.0\pm 0.9$   $\mu\text{M}$  NR, in conditioned medium from cells grown in SDC to an OD of 3 (Table 3). Adding 1 mM NA, increased the extracellular NR produced by both PAB076 and PAB038 to a



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concentration of  $7.7 \pm 1.1$   $\mu$ M and  $3.9 \pm 1.5$   $\mu$ M respectively. Changing the niacin to Nam or supplementing with both niacins did not further improve the NR yield from the PAB076.

TABLE 3

Strain and Condition	[NR] $\mu$ M
Wild-type SDC (OD 3)	$0.12 \pm 0.4$
PAB038 SDC (OD 3)	$1.20 \pm 0.4$
PAB038 SDC + 1 mM NA (OD 3)	$3.90 \pm 1.5$
PAB076 SDC (OD 3)	$4.06 \pm 0.9$
PAB076 SDC + 1 mM NA (OD 3)	$7.70 \pm 1.1$
PAB076 SDC + 1 mM Nam (OD 3)	$7.17 \pm 0.2$
PAB076 SDC + 1 mM Nam & NA (OD 3)	$7.30 \pm 0.3$
PAB076 YPD + 1 mM NA (OD 15)	$10.60 \pm 5.6$
PAB076 2X YPD + 1 mM NA (OD 21)	$21.20 \pm 4.6$
PAB076 SDC + 5 mM NA (OD 7)	$16.80 \pm 0.3$
PAB076 2X SDC + 5 mM NA (OD 13)	$20.80 \pm 4.2$
PAB076 2X YPD + 5 mM NA (OD 60)	$28.15 \pm 8.5$

By adding NA or Nam, the amount of extracellular NR produced could be doubled. To further increase the yield, cells were cultured to extremely high densities. PAB076 was incubated in YPD, 2XYPD, SDC or 2XSDC and growth was measured over a period of 31 hours. Surprisingly, PAB076 was able to grow to an unusually high density in all three media formulations (Table 4). For example, this strain attained an OD of 29 when grown in YPD and an OD of 35 when grown in 2XYPD. To determine the genetic cause of this phenotype, the growth of other related strains was assayed (Table 4). Only one other strain, nrt1 fun26 (PAB75), had this unusual ability to grow to high cell density. The common element present in these two strains and absent in the others was an intact URA3 gene. URA3 was used to knock out nrt1 in the PAB076 strain and fun26 in PAB075. Other nonrelated strains chosen from lab stocks also had the same URA3-dependent high growth phenotype.

TABLE 4

Strain and Condition	OD at 31 hours
nrk1 urh1 pnp1 nrt1 URA3 2X YPD	35.0
nrk1 urh1 pnp1 nrt1 URA3 YPD	29.0
nrk1 urh1 pnp1 nrt1 URA3 SDC	7.0
nrk1 urh1 pnp1 ura3 2X YPD	12.9
nrk1 urh1 pnp1 ura3 YPD	8.1
nrk1 urh1 pnp1 ura3 SDC	6.4
Wild-type (ura3) 2X YPD	12.2
Wild-type (ura3) SDC	5.5
nrt1 ura3 2X YPD	13.0
nrt fun26 URA3 2X YPD	33.7
nrt fun26 URA3 2X YPD	33.2
nrk1 urh1 pnp1 nrt1 URA3 2X YPD	36.1

Growing cells to extremely high cultural density dramatically increased extracellular NR accumulation (Table 3).

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Cells incubated in 2XSDC (5 mM NA) to an OD of 13 and cells incubated in 2XYPD (5 mM NA) to an OD of 60 produced the highest amounts of extracellular NR,  $20.2 \pm 4.3$   $\mu$ M and  $28.1 \pm 8$   $\mu$ M extracellular NR, respectively. Cells that were incubated in 2XYPD, but did not reach stationary phase produced somewhat less extracellular NR than cells grown to an OD of 60. Similarly, cells incubated in 1XSDC or 1XYPD produced significantly less NR than the cells incubated in the 2X formulations. From this data, it appears that the final concentration of NR is both a function final cell number and whether or not the culture reached stationary.

## Example 4

## Purification of NR from PAB076-Conditioned Media

Cultures of PAB076 (500 mL) were grown in 2XSDC or 2XYPD with 5 mM NA, to an OD of 13 and 60, respectively. To extract NR from this medium, a two step process was implemented that first concentrated NR by lyophilization and then separated NR from contaminants using SP-SEPHADEX chromatography. The SP-SEPHADEX fractions were analyzed by HPLC. NA and the majority of the media components eluted in the first 100 ml of the run that contained no salt (FIG. 1). NR was retained by the resin and eluted between 20 and 50 mM NaCl in fractions 27-36. The majority of these fractions were more than 98% pure NR, although the early fractions contained trace amounts of NA. Each fraction was concentrated by lyophilization and the concentration of NR was determined by absorbance at 259 nm. The total yield was  $\sim 700$   $\mu$ g of NR from 150 ml of the media or 5.6 mg/L. Based on MALDI-MS measurements, the concentration of NR in the conditioned 2XYPD prior to extraction was  $\sim 8$  mg/L. It was found that NR from fraction 28 and from pooled fractions 31-34 (added at 10  $\mu$ M) was capable of increasing intracellular NAD<sup>+</sup> in wild-type yeast as efficiently as chemically or enzymatically synthesized NR.

In so far as 2XSDC media could not be effectively fractionated by SP-SEPHADEX because of the high salt content of this media, conditioned 2XSDC medium would require desalting (e.g., with a disposable C18 spin columns) prior to chromatography.

In addition to the above-described approaches, other improvements are contemplated for increasing the yield of NR. These include the use of a chemostat fermenter and the use of industrial scale preparative HPLC chromatography; and genetically engineering a PAB076 strain that also over-expresses the major NMN 5' nucleotidase thereby increasing extracellular NR production and lowering the concentration of NA supplementation. The recommended daily allowance of niacin is 15 mg, and with only slight improvements made possible by industrialization, one liter or less of yeast would be able to produce the daily Niacin requirement in the form of NR.

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&lt;213&gt; ORGANISM: Artificial sequence

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&lt;223&gt; OTHER INFORMATION: Synthetic oligonucleotide

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&lt;211&gt; LENGTH: 84

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60

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84

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&lt;213&gt; ORGANISM: Artificial sequence

&lt;220&gt; FEATURE:

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Ala Thr Leu Ile His Glu Asp Asp Phe Tyr Lys His Asp Asn Glu Val  
 35 40 45

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Pro Val Asp Ala Lys Tyr Asn Ile Gln Asn Trp Asp Ser Pro Glu Ala  
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Leu Asp Phe Lys Leu Phe Gly Lys Glu Leu Asp Val Ile Lys Gln Thr  
65 70 75 80

Gly Lys Ile Ala Thr Lys Leu Ile His Asn Asn Asn Val Asp Asp Pro  
85 90 95

Phe Thr Lys Phe His Ile Asp Arg Gln Val Trp Asp Glu Leu Lys Ala  
100 105 110

Lys Tyr Asp Ser Ile Asn Asp Asp Lys Tyr Glu Val Val Ile Val Asp  
115 120 125

Gly Phe Met Ile Phe Asn Asn Thr Gly Ile Ser Lys Lys Phe Asp Leu  
130 135 140

Lys Ile Leu Val Arg Ala Pro Tyr Glu Val Leu Lys Lys Arg Arg Ala  
145 150 155 160

Ser Arg Lys Gly Tyr Gln Thr Leu Asp Ser Phe Trp Val Asp Pro Pro  
165 170 175

Tyr Tyr Phe Asp Glu Phe Val Tyr Glu Ser Tyr Arg Ala Asn His Ala  
180 185 190

Gln Leu Phe Val Asn Gly Asp Val Glu Gly Leu Leu Asp Pro Arg Lys  
195 200 205

Ser Lys Asn Ile Lys Glu Phe Ile Asn Asp Asp Asp Thr Pro Ile Ala  
210 215 220

Lys Pro Leu Ser Trp Val Cys Gln Glu Ile Leu Lys Leu Cys Lys Asp  
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&lt;213&gt; ORGANISM: Schizosaccharomyces pombe

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Gly Lys Ser Thr Leu Cys Gln Leu Leu His Ala Ile Phe Glu Gly Ser  
20 25 30

Ser Leu Val His Glu Asp Asp Phe Tyr Lys Thr Asp Ala Glu Ile Pro  
35 40 45

Val Lys Asn Gly Ile Ala Asp Trp Asp Cys Gln Glu Ser Leu Asn Leu  
50 55 60

Asp Ala Phe Leu Glu Asn Leu His Tyr Ile Arg Asp His Gly Val Leu  
65 70 75 80

Pro Thr His Leu Arg Asn Arg Glu Asn Lys Asn Val Ala Pro Glu Ala  
85 90 95

Leu Ile Glu Tyr Ala Asp Ile Ile Lys Glu Phe Lys Ala Pro Ala Ile  
100 105 110

Pro Thr Leu Glu Gln His Leu Val Phe Val Asp Gly Phe Met Met Tyr  
115 120 125

Val Asn Glu Asp Leu Ile Asn Ala Phe Asp Ile Arg Leu Met Leu Val  
130 135 140

Thr Asp Phe Asp Thr Leu Lys Arg Arg Arg Glu Ala Arg Thr Gly Tyr  
145 150 155 160

Ile Thr Leu Glu Gly Phe Trp Gln Asp Pro Pro His Tyr Phe Glu Asn  
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Tyr Val Trp Pro Gly Tyr Val His Gly His Ser His Leu Phe Val Asn  
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Gly Asp Val Thr Gly Lys Leu Leu Asp Lys Arg Ile Gln Leu Ser Pro  
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Ser Ser Lys Met Ser Val Arg Asp Asn Val Gln Trp Ala Ile Asn Ser  
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Ser Ser Val Leu His Glu Asp Asp Phe Tyr Lys Pro Asp Ala Gln Ile  
35 40 45

Pro Leu Asn Glu Lys Tyr Gln Ile Ala Asp Trp Asp Cys Pro Glu Ala  
50 55 60

Leu Asp Ile Pro Ala Phe Lys Arg Glu Leu Asp Gln Ile Lys Glu Thr  
65 70 75 80

Gly Leu Ile Lys Ser Lys Leu Ile His Asn Asp Asn Val Asp Asp Ile  
85 90 95

Thr Lys Phe Asp Ile Ser Pro Glu Asp Trp Asp Ser Leu Lys Arg Lys  
100 105 110

Tyr Ala Ile Val Gln Asn Ser Asp Leu Lys Ile Val Leu Val Asp Gly  
115 120 125

Phe Met Ile Phe Asn Asp Glu Glu Leu Thr Lys Lys Phe Asp Ile Lys  
130 135 140

Ile Phe Val Arg Ala Pro Tyr Glu Val Leu Lys Lys Arg Arg Asn Ala  
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Arg Ala Gly Tyr Lys Thr Ile Asp Ser Tyr Trp Val Asp Pro Pro Tyr  
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Tyr Phe Asp Glu Phe Val Tyr Lys Ser Tyr Arg Asn Glu His Lys Tyr  
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Met Phe Glu Asp Glu Asp Ile Glu Gly Gln Leu Lys Arg Asn Thr Gly  
195 200 205

Leu Phe Glu Leu Lys Asn Asp Asp Asp Ile Asn Ile Ser Asp Ala Leu  
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Ala Leu Ser Tyr Ile Arg Asp His Gly Gln Leu Pro Pro Arg Leu Lys		
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Ser Ile Gln Asp Leu Asn Glu Lys Ser Asp Ser Gly Val Asp Glu Gly		
	100	105 110
Thr Ile Leu Gln Leu Gln Gln Glu Val Gly Gly Arg Leu Arg Ala Arg		
	115	120 125
Ala Pro Ala Lys Arg Thr Ile Ala Phe Leu Glu Gly Phe Leu Leu Tyr		
	130	135 140
Ser Pro Pro Glu Ser Glu Asp Lys Asp His Val Leu Arg Ser Val His		
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Lys Asn Ile Asp Val His Leu Phe Leu Pro Ala Pro Tyr Asp Met Val		
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Lys Ser Arg Arg Glu Gly Arg Ser Gly Tyr Val Thr Ser Gly Pro Ala		
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Pro Glu Pro Thr Ser Leu Pro Gln Arg Ser Ser Val Ser Asp Glu Val		
	195	200 205
Asp Leu Glu Gly Glu Asp Asp Arg Pro Pro Gln Asn Phe Trp Thr Asp		
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Pro Pro Gly Tyr Val Asp Asp Ile Val Trp Pro Arg Tyr Val Gln Asp		
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His Ala Trp Leu Ile Leu Pro Glu Gly Glu Ser Gln Lys Ser Asn Thr		
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Leu Ser Ala Asp Ser Gln Glu Leu Val Asn Lys Val Gly Gln Gly Val		
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Asn Leu Arg Thr Asn Ala Gly Val Ile Val Ala Pro Gly Glu Gly Thr		
	275	280 285
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Ile Lys Asn Ser Arg Leu Ile His Leu Asp Asp Phe Tyr Leu Ala Asp		
	35	40 45
His Leu Ile Pro Val Asp Pro Val Thr Gly Gln Gln Asn Trp Asp Val		
	50	55 60
Pro Glu Ala Leu Asp Phe Ala Arg Phe Thr Ser Tyr Ile Lys Ser Ile		
	65	70 75 80
Arg Gln Ser His Asn Leu Glu Asp Lys Ile Asp Thr Leu Glu Pro Asp		
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Thr Asn Leu Lys Leu Thr Ala Gln Glu Val Gln Gln Phe Glu Ala Lys		

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Val Lys Leu Phe Phe His Ala Ser Phe Glu Thr Leu Lys Asn Arg Arg		
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Glu Ser Arg Lys Gly Tyr Asn Thr Val Glu Gly Phe Trp Val Asp Pro		
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Pro Asn Tyr Phe Arg Asp Met Val Trp Pro Ala Tyr Glu Ser Ser His		
180	185	190
Lys Tyr Leu Phe Glu Asn Lys Asp Val Asp Gly Val Leu Lys Ser Glu		
195	200	205
Tyr Lys Thr Gln Tyr Gln Ile His Asp Ile Arg Asn Glu Thr Gly Val		
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Lys Leu Tyr Glu Val Val Asp Trp Ser Leu Gln His Ile Phe Ala Met		
225	230	235
Val Lys Arg Leu		

&lt;210&gt; SEQ ID NO 18

&lt;211&gt; LENGTH: 340

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Saccharomyces cerevisiae*

&lt;400&gt; SEQUENCE: 18

Met Thr Val Ser Lys Ile Pro Ile Trp Leu Asp Cys Asp Pro Gly His		
1	5	10
Asp Asp Ala Ile Ala Ile Leu Leu Gly Cys Phe His Pro Ala Phe Asn		
20	25	30
Leu Leu Gly Ile Ser Thr Cys Phe Gly Asn Ala Pro Pro Glu Asn Thr		
35	40	45
Asp Tyr Asn Ala Arg Ser Leu Leu Thr Ala Met Gly Lys Ala Gln Ala		
50	55	60
Ile Pro Val Tyr Lys Gly Ala Gln Arg Pro Trp Lys Arg Glu Pro His		
65	70	75
Tyr Ala Pro Asp Ile His Gly Ile Ser Gly Leu Asp Gly Thr Ser Leu		
85	90	95
Leu Pro Lys Pro Thr Phe Glu Ala Arg Thr Asp Lys Thr Tyr Ile Glu		
100	105	110
Ala Ile Glu Glu Ala Ile Leu Ala Asn Asn Gly Glu Ile Ser Phe Val		
115	120	125
Ser Thr Gly Ala Leu Thr Thr Leu Ala Thr Val Phe Arg Cys Lys Pro		
130	135	140
Tyr Leu Lys Lys Ser Val Lys Tyr Ile Ser Ile Met Gly Gly Gly Leu		
145	150	155
His Gly Leu Gly Asn Cys Asn Pro Asn Leu Ser Ala Glu Phe Asn Val		
165	170	175
Trp Ile Asp Pro Asp Ala Ala Asn Tyr Ile Phe Arg Asp Pro Asp Val		
180	185	190
Lys Asp Lys Cys Ile Val Val Pro Leu Asn Leu Thr His Lys Ala Ile		
195	200	205
Ala Thr Tyr Lys Val Asn Glu Met Ile Tyr Asn Glu Lys Asn Asn Ser		
210	215	220
Lys Leu Arg Glu Leu Phe Leu Glu Leu Phe Gln Phe Phe Ala His Thr		
225	230	235
		240

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Tyr Lys Asp Met Gln Gly Phe Glu Ser Gly Pro Pro Ile His Asp Pro  
 245 250 255  
 Val Ala Leu Met Pro Leu Leu Glu Phe Tyr Gly Trp Asp Pro Ser Ser  
 260 265 270  
 Ala Val Gly Phe Arg Tyr Lys Arg Met Asp Ile Ser Cys Ile Asp Asp  
 275 280 285  
 Val Phe Asn Glu Asn Ser Gly Lys Ile Ile Ile Glu Lys Glu Tyr Pro  
 290 295 300  
 Asn Asp Ser Asp Val Gly Thr Ile Ile Gly Leu Asp Leu Asn Ile Gln  
 305 310 315 320  
 Tyr Phe Trp Asp Gln Ile Phe Glu Ala Leu Asn Arg Ala Asp Lys Met  
 325 330 335  
 Ser Thr Ile Gly  
 340

<210> SEQ ID NO 19  
 <211> LENGTH: 330  
 <212> TYPE: PRT  
 <213> ORGANISM: Schizosaccharomyces pombe

<400> SEQUENCE: 19

Met Thr Asn Thr Ile Asp Ser Phe Gln Lys Gly Ser Ala Leu Glu Asn  
 1 5 10 15  
 Tyr Asn Ile Trp Ile Asp Cys Asp Pro Gly His Asp Asp Val Val Ala  
 20 25 30  
 Leu Thr Leu Ala Ala Cys Ala Gly His Cys Lys Ile Leu Gly Val Ser  
 35 40 45  
 Thr Val His Gly Asn Thr Thr Leu Glu Phe Thr Thr Lys Asn Ala Leu  
 50 55 60  
 Ala Val Met Glu Leu Leu Asn Gln Asp Val Asp Val His Ala Gly Ala  
 65 70 75 80  
 Ala Lys Pro Leu Met Arg Glu Ser Ala Phe Ala Thr His Ile His Gly  
 85 90 95  
 Thr Asn Gly Leu Ala Gly Ile Ser Leu Leu Pro Asp Tyr Pro Lys Lys  
 100 105 110  
 Lys Ala Thr Pro Asp Ala Val Phe Ala Met Tyr Thr Thr Ile Ser Asn  
 115 120 125  
 Tyr Pro Glu Pro Val Thr Leu Val Ala Thr Gly Pro Leu Thr Asn Ile  
 130 135 140  
 Ala Leu Leu Leu Ala Thr Tyr Pro Ser Val Thr Asp Asn Ile Glu Arg  
 145 150 155 160  
 Phe Ile Phe Met Gly Gly Ser Thr Gly Ile Gly Asn Ile Thr Ser Gln  
 165 170 175  
 Ala Glu Phe Asn Val Tyr Ala Asp Pro Glu Ala Ala Arg Leu Val Leu  
 180 185 190  
 Glu Thr Lys Ser Leu Ile Gly Lys Leu Phe Met Val Pro Leu Asp Val  
 195 200 205  
 Thr His Lys Val Leu Leu Asp Ala Asn Ile Ile Gln Leu Leu Arg Gln  
 210 215 220  
 His Ser Asn Pro Phe Ser Ser Thr Leu Val Glu Leu Met Thr Val Phe  
 225 230 235 240  
 Gln Gln Thr Tyr Glu Asn Val Tyr Gly Ile Arg Asn Gly Val Pro Val  
 245 250 255  
 His Asp Val Cys Ala Val Ala Leu Ala Leu Trp Pro Ser Leu Trp Thr  
 260 265 270

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Ser Arg Ser Met Tyr Val Thr Val Ser Leu Asp Ser Leu Thr Leu Gly  
 275 280 285

Arg Thr Val Cys Asp Val Trp Ser Gln Gln Asn Gln Tyr Pro Ala Asn  
 290 295 300

Val His Val Val Leu Glu Ala Asp Val Ser Leu Phe Trp Glu Thr Phe  
 305 310 315 320

Ile Gly Val Ile Asp Arg Leu Asn Tyr Leu  
 325 330

<210> SEQ ID NO 20  
 <211> LENGTH: 338  
 <212> TYPE: PRT  
 <213> ORGANISM: Kluyveromyces lactis  
 <400> SEQUENCE: 20

Met Thr Gly Asn Ser Val Ile Pro Ile Trp Val Asp Cys Asp Pro Gly  
 1 5 10 15

His Asp Asp Ala Val Ala Ile Leu Leu Ser Cys Phe His Pro Ser Ile  
 20 25 30

Arg Leu Leu Gly Ile Ser Ala Ser Tyr Gly Asn Ala Ser Pro Glu Asn  
 35 40 45

Thr Leu Tyr Asn Thr Leu Ser Leu Leu Thr Ala Phe Gly Lys Gln Asp  
 50 55 60

Glu Val Pro Val Tyr Lys Gly Ala Gln Arg Pro Trp Val Arg Asp Val  
 65 70 75 80

Ala Tyr Ala Pro Asp Ile His Gly Glu Thr Gly Leu Asp Gly Thr Thr  
 85 90 95

Leu Leu Pro Lys Pro Lys Arg Ser Phe Val Asp Ala Asp Tyr Ile Lys  
 100 105 110

Ala Met Glu Asn Ala Ile Leu Ala Asn Gly Gly Asn Ile Ala Leu Val  
 115 120 125

Ser Thr Gly Thr Leu Thr Ser Ile Ala Thr Leu Phe Lys Glu Lys Pro  
 130 135 140

Tyr Leu Lys Glu Gln Val Arg Tyr Ile Ser Ile Met Gly Gly Gly Leu  
 145 150 155 160

His Ala Gly Asn Arg Asn Asp Asn Asp Ser Ala Glu Phe Asn Ile Trp  
 165 170 175

Ala Asp Pro Asp Ala Ala Asp Phe Ile Leu Asn Asp Glu Asp Ile Lys  
 180 185 190

His Lys Cys Ile Leu Ser Pro Leu Asp Leu Thr His Thr Cys Ile Ala  
 195 200 205

Thr Glu Tyr Ile Asp Lys Thr Ile Leu Gly Asp Gly Ser Cys Lys Leu  
 210 215 220

Arg Lys Leu Phe Tyr Glu Leu Phe Leu Phe Phe Ala Lys Thr Tyr Lys  
 225 230 235 240

Asn Lys Gln Gly Phe Glu Ala Gly Pro Pro Val His Asp Pro Val Thr  
 245 250 255

Leu Met Pro Leu Leu Tyr Leu Tyr Gly His Ile Ser Asn Asp Ile Leu  
 260 265 270

Arg Phe Lys Tyr Gly Arg Phe Asp Leu Ser Ile Asp Lys Asn Gln Asp  
 275 280 285

Ser Ile Asn Tyr Gly Arg Thr Ile Val Thr Gln Glu Tyr Pro Ser Asp  
 290 295 300

Ser Asn Phe Gly Leu Met Val Gly Leu Gln Ile Asn Val Asp Phe Phe  
 305 310 315 320



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Trp Asn Gln Val Leu Asn Ala Ile Asp Val Ala Glu Asn Tyr Pro Gly  
 325 330 335

Ser Leu

<210> SEQ ID NO 21  
 <211> LENGTH: 375  
 <212> TYPE: PRT  
 <213> ORGANISM: *Aspergillus oryzae*

<400> SEQUENCE: 21

Met His Ser Ser Ser Asp Ile Pro Ile Pro Leu Trp Leu Asp Cys Asp  
 1 5 10 15  
 Pro Gly His Asp Asp Ala Phe Ala Ile Leu Leu Ala Ala His His Pro  
 20 25 30  
 Ser Leu Asn Leu Leu Gly Ile Thr Thr Val His Gly Asn Ala Ser Leu  
 35 40 45  
 Glu Asn Thr Thr Asn Asn Ala Thr Arg Ile Leu Glu Ala Ile Gly Arg  
 50 55 60  
 Pro Glu Ile Pro Val Tyr Pro Gly His Lys Lys Pro Phe Cys Arg Pro  
 65 70 75 80  
 Ala Ile His Ala Pro Asn Ile His Gly Asp Ser Gly Ile Asp Gly Thr  
 85 90 95  
 Glu Leu Leu Pro Lys Ala Thr Lys Ser Pro Ile Thr Asp Lys Asn Pro  
 100 105 110  
 Ile Leu Ala Met Arg Asp Ala Leu Leu Ala Gln Pro Lys Gly Thr Pro  
 115 120 125  
 Trp Val Ile Ala Thr Gly Thr Leu Thr Asn Val Ala Leu Leu Phe Ala  
 130 135 140  
 Thr Phe Pro Glu Val Ala Glu His Ile Gln Gly Leu Ser Ile Met Gly  
 145 150 155 160  
 Gly Gly Val Gly Gly Gly Phe Thr Asp Ala Pro Met Ser Arg Leu Val  
 165 170 175  
 Gly Glu Glu Ser Arg Ile Gly Asn Ile Thr Pro Leu Ala Glu Phe Asn  
 180 185 190  
 Ile Tyr Cys Asp Pro Glu Ala Ser Gln Ser Ile Phe Ser Asn Pro Val  
 195 200 205  
 Leu Ala Ser Lys Thr Thr Leu Ile Thr Leu Asp Leu Thr His Gln Val  
 210 215 220  
 Leu Ala Ser His Ser Val Gln Ser Arg Val Leu His Gly Gly Asp Asp  
 225 230 235 240  
 Leu Ser Val Pro Pro Thr Val Leu Arg Gln Met Leu Phe Asp Leu Leu  
 245 250 255  
 Val Phe Phe Ala Ser Thr Tyr Glu Asn Val Phe Gly Leu Thr Ser Gly  
 260 265 270  
 Pro Pro Leu His Asp Pro Leu Ala Val Ala Val Ile Leu Ser Thr Leu  
 275 280 285  
 Asn Pro Glu Tyr Ala Lys Arg His Pro Asp Gln Val Leu Lys Phe Asp  
 290 295 300  
 Asp Arg Asn Gly Glu Arg Phe Asp Val Asp Val Val Thr Asp Gly Leu  
 305 310 315 320  
 His Gly Thr Asp Val Glu Leu Val Gly Glu Leu Gly Arg Ser Lys Val  
 325 330 335  
 Ile Ser Gly Thr Thr Gly Val Ala Ile Pro Arg Gly Val Asp Leu Asp  
 340 345 350

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Ala Phe Trp Asn Met Ile Leu Asp Cys Leu Arg Arg Ala Asp Glu Cys  
 355 360 365

Asn Ala Ala Arg Lys Leu Ala  
 370 375

<210> SEQ ID NO 22  
 <211> LENGTH: 348  
 <212> TYPE: PRT  
 <213> ORGANISM: *Pichia stipitis*

<400> SEQUENCE: 22

Met Thr Val Gly Glu Lys Ile Pro Ile Trp Leu Asp Cys Asp Pro Gly  
 1 5 10 15  
 Asn Asp Asp Ala Phe Ala Ile Leu Leu Ala Leu Phe Asp Pro Arg Phe  
 20 25 30  
 Glu Leu Leu Gly Ile Ser Thr Val His Gly Asn Ala Pro Leu Ser Tyr  
 35 40 45  
 Thr Thr His Asn Ala Leu Ser Leu Leu Asp Ser Leu Gly Val Glu Pro  
 50 55 60  
 Gly Thr Val Lys Val Tyr Ala Gly Ser Glu Thr Pro Leu Val Asn Ala  
 65 70 75 80  
 Pro Gln Ser Ala Pro Glu Ile His Gly Thr Thr Gly Ile Gly Gly Val  
 85 90 95  
 Glu Phe Pro Glu Val Thr Lys Asn Lys Val Ala Thr Asp Val Gly Tyr  
 100 105 110  
 Leu Glu Ala Met Lys Gln Ala Ile Leu Ser His Glu Asn Glu Leu Cys  
 115 120 125  
 Leu Val Cys Thr Gly Thr Leu Thr Asn Val Ser Lys Leu Ile Thr Glu  
 130 135 140  
 Cys Pro Ala Ile Ile Pro Lys Ile Arg Tyr Val Ser Ile Met Gly Gly  
 145 150 155 160  
 Ala Phe Asn Leu Gly Asn Val Thr Pro Tyr Ala Glu Phe Asn Phe Tyr  
 165 170 175  
 Ala Asp Pro His Ala Ala Lys His Val Leu Ala Glu Leu Gly Pro Lys  
 180 185 190  
 Ile Ile Leu Ser Pro Leu Asn Ile Thr His Lys Ala Thr Ala Thr Glu  
 195 200 205  
 Ser Ile Arg Asn Gln Met Tyr Asp Ser Glu Asp Pro His Arg Asn Ser  
 210 215 220  
 Asp Ile Arg Asn Met Phe Tyr Ser Ile Leu Met Phe Phe Ser His Ser  
 225 230 235 240  
 Tyr Ile Lys Lys Tyr Gly Ile Thr Glu Gly Pro Pro Val His Asp Pro  
 245 250 255  
 Leu Ala Leu Tyr Cys Leu Leu Pro Phe Leu Gln Gln Asp Lys Asp Tyr  
 260 265 270  
 Lys Tyr Lys Tyr Leu Arg Arg Lys Val Ser Val Ile Thr Glu Gly Glu  
 275 280 285  
 His Ser Gly Glu Ser Ile Leu Leu Asn Gly Asn Ser Asp Ser Ser Val  
 290 295 300  
 Glu Glu Glu Asp Gly Val Tyr Ile Gly Gln Asp Ile Asp Val Asp Gln  
 305 310 315 320  
 Phe Trp Arg Thr Val Leu Arg Ala Val Asn Val Ala Asp Val Thr Ile  
 325 330 335  
 Lys Gln Glu Ile Asn Gly Ala Gln Lys Val Met Val  
 340 345

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<210> SEQ ID NO 23  
 <211> LENGTH: 311  
 <212> TYPE: PRT  
 <213> ORGANISM: *Saccharomyces cerevisiae*

<400> SEQUENCE: 23

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Met Ser Asp Ile Leu Asn Val Ser Gln Gln Arg Glu Ala Ile Thr Lys
1           5           10           15
Ala Ala Ala Tyr Ile Ser Ala Ile Leu Glu Pro His Phe Lys Asn Thr
20           25           30
Thr Asn Phe Glu Pro Pro Arg Thr Leu Ile Ile Cys Gly Ser Gly Leu
35           40           45
Gly Gly Ile Ser Thr Lys Leu Ser Arg Asp Asn Pro Pro Val Thr
50           55           60
Val Pro Tyr Gln Asp Ile Pro Gly Phe Lys Lys Ser Thr Val Pro Gly
65           70           75           80
His Ser Gly Thr Leu Met Phe Gly Ser Met Asn Gly Ser Pro Val Val
85           90           95
Leu Met Asn Gly Arg Leu His Gly Tyr Glu Gly Asn Thr Leu Phe Glu
100          105          110
Thr Thr Phe Pro Ile Arg Val Leu Asn His Met Gly His Val Arg Asn
115          120          125
Leu Ile Val Thr Asn Ala Ala Gly Gly Ile Asn Ala Lys Tyr Gln Ala
130          135          140
Cys Asp Leu Met Cys Ile Tyr Asp His Leu Asn Ile Pro Gly Leu Ala
145          150          155          160
Gly Gln His Pro Leu Arg Gly Pro Asn Leu Asp Glu Asp Gly Pro Arg
165          170          175
Phe Leu Ala Leu Ser Asp Ala Tyr Asp Leu Glu Leu Arg Lys Leu Leu
180          185          190
Phe Lys Lys Trp Lys Glu Leu Lys Ile Gln Arg Pro Leu His Glu Gly
195          200          205
Thr Tyr Thr Phe Val Ser Gly Pro Thr Phe Glu Thr Arg Ala Glu Ser
210          215          220
Lys Met Ile Arg Met Leu Gly Gly Asp Ala Val Gly Met Ser Thr Val
225          230          235          240
Pro Glu Val Ile Val Ala Arg His Cys Gly Trp Arg Val Leu Ala Leu
245          250          255
Ser Leu Ile Thr Asn Thr Cys Val Val Asp Ser Pro Ala Ser Ala Leu
260          265          270
Asp Glu Ser Pro Val Pro Leu Glu Lys Gly Lys Ala Thr His Ala Glu
275          280          285
Val Leu Glu Asn Gly Lys Ile Ala Ser Asn Asp Val Gln Asn Leu Ile
290          295          300
Ala Ala Val Met Gly Glu Leu
305          310

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<210> SEQ ID NO 24  
 <211> LENGTH: 315  
 <212> TYPE: PRT  
 <213> ORGANISM: *Schizosaccharomyces pombe*

<400> SEQUENCE: 24

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Met Thr Ala Thr Ser Phe Leu His Gln Ala Lys Gln Gln Pro His His
1           5           10           15
Thr Glu Pro Tyr Ile Lys Ala Leu Glu Ala Arg Glu Tyr Ile Ile Glu

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20	25	30
Gln Val Pro Glu Glu Leu Ser Lys	Pro Lys Val Ala Ile Ile Cys Gly	
35	40	45
Ser Gly Leu Gly Thr Leu Ala Ser Gly	Leu Ser Ala Pro Val Tyr Glu	
50	55	60
Val Pro Tyr Glu Asp Ile Pro His Phe	His Val Ser His Val Pro Gly	
65	70	75
His Ala Ser Lys Leu Tyr Phe Ala Phe	Leu Gly Glu Lys Arg Val Pro	
85	90	95
Thr Met Ile Leu Ala Gly Arg Tyr His	Ser Tyr Glu Gly Tyr Pro Ile	
100	105	110
Glu Ala Thr Thr Phe Pro Val Arg Leu	Met Lys Val Met Gly Val Glu	
115	120	125
Val Met Val Val Thr Asn Ala Ala Gly	Gly Leu Asn Gln Gly Phe Lys	
130	135	140
Val Gly Asp Leu Met Ile Leu Lys Asp	His Ile Asn Phe Pro Gly Leu	
145	150	155
Ala Gly Met Asn Pro Leu Arg Gly Pro	Asn Ala His Glu Phe Gly Val	
165	170	175
Arg Phe Pro Pro Leu Ser Asp Ala Tyr	Asp Leu Glu Leu Arg Lys Leu	
180	185	190
Val Tyr Asp Ala Ala Lys Ala His Lys	Val Ser Arg Thr Ile His Glu	
195	200	205
Gly Cys Tyr Ala Phe Val Ser Gly Pro	Cys Phe Glu Thr Arg Ala Glu	
210	215	220
Ser Arg Met Leu Ala Leu Met Gly Ala	Asp Cys Val Gly Met Ser Thr	
225	230	235
Val Pro Glu Val Val Val Ala Arg His	Cys Gly Ile Arg Val Leu Ala	
245	250	255
Ile Ser Leu Val Thr Asn Asn Val Val	Val Glu Glu Ser Pro Ser Ala	
260	265	270
Lys Asp Leu Val Glu Val Asp Ser Asn	Val Met Ser Lys Gly Ala Ala	
275	280	285
Asn His Leu Glu Val Leu Glu Val Gly	Ile Ala Ala Ala Asp Val	
290	295	300
Arg Thr Met Val Glu Thr Ile Val Asn	Phe Ile	
305	310	315

&lt;210&gt; SEQ ID NO 25

&lt;211&gt; LENGTH: 306

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Kluyveromyces lactis

&lt;400&gt; SEQUENCE: 25

Met Ser Ser Leu Asp Ile Asn Glu Gln	Arg Ala Leu Ile Lys Ser Ala
1	15
His Arg Tyr Ile Ser Glu Lys Leu Glu	Asp His Phe Ser Ser Glu Phe
20	30
Leu Pro Lys Ala Leu Val Ile Cys Gly	Ser Gly Leu Ser Gly Ile Ser
35	45
Thr Lys Ile Ala Asp Glu Pro Lys Pro	Leu Ile Leu Ser Tyr Ser Thr
50	60
Ile Pro Gly Phe Lys Val Ser Thr Val	Pro Gly His Ser Gly Glu Leu
65	80
Ile Phe Gly Tyr Met Asn Gly Ala Pro	Val Val Leu Met Asn Gly Arg



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85	90	95
Leu His Ser Tyr Glu Gly His Ser Leu Ala Glu Thr Val His Pro Ile		
100	105	110
Arg Ala Leu His Leu Leu Gly Ser Ile Asn Val Leu Ile Val Thr Asn		
115	120	125
Ala Ala Gly Gly Ile Asn Ala Ser Phe Lys Ala Gly Asp Leu Met Cys		
130	135	140
Val Tyr Asp His Ile Asn Phe Pro Gly Leu Cys Gly Phe His Pro Leu		
145	150	155
Arg Gly Ala Asn Phe Asp Glu Phe Gly Pro Arg Phe Leu Ala Thr Ser		
165	170	175
Asp Ala Tyr Asp Leu Glu Leu Arg Lys Leu Leu Phe Ser Lys Lys Lys		
180	185	190
Glu Leu Asn Ile Glu Arg Lys Ile His Glu Gly Thr Tyr Ser Tyr Val		
195	200	205
His Gly Pro Thr Phe Glu Ser Arg Ala Glu Ser Arg Phe Leu Arg Leu		
210	215	220
Ala Gly Thr Asp Ala Val Gly Met Ser Thr Val Pro Glu Val Val Thr		
225	230	235
Ala Arg His Cys Gly Trp Arg Val Leu Ala Leu Ser Leu Ile Thr Asn		
245	250	255
Glu Cys Val Val Asp Pro Pro Ala Ser Ala His Asp Glu Asn Pro Val		
260	265	270
Pro Ile Gln Glu Gly Lys Ala Thr His Glu Glu Val Leu Glu Asn Ser		
275	280	285
Ala Lys Ala Ser Lys Asp Val Gln Glu Leu Ile Phe Ser Val Val Ala		
290	295	300
Glu Ile		
305		

&lt;210&gt; SEQ ID NO 26

&lt;211&gt; LENGTH: 598

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Saccharomyces cerevisiae

&lt;400&gt; SEQUENCE: 26

Met Ser Phe Ser Ser Ile Val Ser Lys Phe Leu Arg Tyr Leu Glu Ile		
1	5	10
Pro Ala Lys Asn Arg Thr Ala Val Asn Phe Leu Arg Asn Pro Asp Leu		
20	25	30
Gln Pro Ile Lys Ser Ala Asn Gln Thr Trp Gly Phe Trp Ser Asn Leu		
35	40	45
Ala Tyr Trp Gly Ala Val Ser Phe Thr Ala Gly Thr Trp Met Ser Gly		
50	55	60
Ser Ala Ala Leu Ser Val Gly Leu Ser Tyr Pro Glu Thr Ile Val Ser		
65	70	75
Phe Leu Leu Gly Asn Val Leu Thr Ile Ile Phe Thr Met Ala Asn Ser		
85	90	95
Tyr Pro Gly Tyr Asp Trp Lys Ile Gly Phe Thr Leu Ala Gln Arg Phe		
100	105	110
Val Phe Gly Ile Tyr Gly Ser Ala Phe Gly Ile Ile Ile Arg Ile Leu		
115	120	125
Met Ser Ile Val Asn Tyr Gly Ser Asn Ala Trp Leu Gly Gly Leu Ser		
130	135	140
Ile Asn Met Ile Leu Asp Ser Trp Ser His His Tyr Leu His Leu Pro		

-continued

145	150	155	160
Asn Thr Leu Ser Pro Ser Val Ala Met Thr Thr Lys Gln Leu Val Gly			
	165	170	175
Phe Ile Ile Phe His Val Leu Thr Ala Leu Cys Tyr Phe Met Lys Pro			
	180	185	190
Tyr His Met Asn Tyr Leu Leu Ile Trp Ser Cys Val Ala Thr Cys Phe			
	195	200	205
Ala Met Leu Gly Ile Val Ile Tyr Leu Thr Lys Asn Ala His Gly Val			
	210	215	220
Gly Glu Leu Phe Thr Ser Thr Lys Ser Thr Val Thr Gly Ser Lys Arg			
	225	230	235
Ala Trp Ala Trp Val Tyr Met Ile Ser Tyr Trp Phe Gly Ser Ile Ser			
	245	250	255
Pro Gly Ser Thr Asn Gln Ser Asp Tyr Ser Arg Phe Gly Ser Ser Asn			
	260	265	270
Leu Ala Ile Trp Thr Gly Ser Val Cys Ala Leu Leu Ile Pro Ala Thr			
	275	280	285
Leu Val Pro Ile Phe Gly Val Ile Ser Ala Ser Thr Cys Asp Lys Leu			
	290	295	300
Tyr Gly Lys Gln Phe Trp Met Pro Met Asp Ile Phe Asp Tyr Trp Leu			
	305	310	315
Thr Asn Asn Tyr Ser Ala Gly Ala Arg Ala Gly Ala Phe Phe Cys Gly			
	325	330	335
Leu Cys Phe Thr Met Ser Gln Met Ser Ser Thr Ile Ser Asn Cys Gly			
	340	345	350
Phe Ala Thr Gly Met Asp Met Ala Gly Leu Leu Pro Lys Tyr Val Asp			
	355	360	365
Ile Lys Arg Gly Ala Leu Phe Cys Ala Cys Ile Ser Trp Ala Cys Leu			
	370	375	380
Pro Trp Asn Phe Tyr Asn Ser Ser Ser Thr Phe Leu Thr Val Met Ser			
	385	390	395
Ser Phe Gly Val Val Met Thr Pro Ile Ile Ala Val Met Ile Cys Asp			
	405	410	415
Asn Phe Leu Ile Arg Lys Arg Gln Tyr Ser Ile Thr Asn Ala Phe Ile			
	420	425	430
Leu Lys Gly Glu Tyr Tyr Phe Thr Lys Gly Val Asn Trp Arg Ala Ile			
	435	440	445
Val Ala Trp Val Cys Gly Met Ala Pro Gly Leu Pro Gly Ile Ala Trp			
	450	455	460
Glu Val Asn Asn Asn Tyr Phe His Asp Ser Gly Ile Val Lys Phe Phe			
	465	470	475
Tyr Gly Asp Ser Phe Phe Ser Phe Leu Ile Ser Phe Phe Val Tyr Trp			
	485	490	495
Gly Leu Cys Val Phe Phe Pro Phe Lys Ile Thr Val Arg His Asp Asp			
	500	505	510
Lys Asp Tyr Tyr Gly Ala Phe Thr Asp Glu Glu Ala Arg Lys Lys Gly			
	515	520	525
Met Ile Pro Tyr Ser Glu Ile Ser Glu Glu Glu Ile Arg Ala Tyr Thr			
	530	535	540
Leu Gly Glu Cys Tyr Thr Thr Gly His Glu Tyr Lys Pro Glu Ser Ser			
	545	550	555
Asp Asn Glu Ser Pro Glu Leu Ile Lys Thr Ser Ser Glu Asn Thr Asn			
	565	570	575

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Val Phe Glu Ile Val His Gln Lys Asp Asp Glu Lys His Ser Phe Ser  
 580 585 590

Thr Thr Gln Gln Val Val  
 595

<210> SEQ ID NO 27  
 <211> LENGTH: 559  
 <212> TYPE: PRT  
 <213> ORGANISM: Schizosaccharomyces pombe

<400> SEQUENCE: 27

Met Glu Asp Pro Lys Ser Asp Glu Lys Phe Asp Ile Gly Ile Ser Glu  
 1 5 10 15  
 Lys Asn Leu Asp Val Gly Phe Gly Glu Ser Ser Ser Val Asp Val Pro  
 20 25 30  
 Val Lys Gly Arg Phe Ala Ser Phe Leu Lys Lys Leu Glu Leu Ser Ser  
 35 40 45  
 Gly Pro Glu Lys Glu Asn Ile Asp Leu Arg Pro Thr Pro Pro Asp Arg  
 50 55 60  
 Arg His Tyr Ser Ala Leu Asp Ile Ile Tyr Leu Trp Ser Cys Asn Gly  
 65 70 75 80  
 Ile Ser Ala Ser Ala Phe Arg Thr Gly Thr Ser Tyr Met Glu Met Gly  
 85 90 95  
 Leu Ser Pro Lys Gln Ala Leu Ala Ala Leu Ile Ala Gly Asn Val Phe  
 100 105 110  
 Ile Ala Met Pro Met Thr Leu Asn Gly Leu Phe Gly Ser His Tyr His  
 115 120 125  
 Ile Pro Phe Ala Val Gln Ser Arg Ala Ser Phe Gly Tyr Tyr Phe Asn  
 130 135 140  
 Thr Leu Ile Ile Leu Leu Arg Phe Ile Ala Gly Leu Phe Tyr Tyr Gly  
 145 150 155 160  
 Thr Asn Val Tyr Thr Gly Ala Glu Cys Val Gln Thr Ile Leu Tyr Ala  
 165 170 175  
 Ile Phe Lys Ser Phe Arg Ser Tyr Lys Asn Arg Leu Pro Ala Asp Ala  
 180 185 190  
 Gly Ile Thr Ser Asp Phe Leu Ile Ser Tyr Phe Val Tyr Trp Val Ile  
 195 200 205  
 Ser Phe Pro Phe His Leu Ile Arg Pro Glu Tyr Leu Gln Arg Phe Phe  
 210 215 220  
 Leu Ile Lys Ser Ile Ser Thr Tyr Ile Ala Cys Phe Ala Met Leu Ile  
 225 230 235 240  
 Phe Leu Leu Cys Asn Val Gly Ser His Val Val Trp Asp Gln Pro Ala  
 245 250 255  
 Thr Val Ser Gly Arg Ser Trp Ser Trp Val Phe Met Cys Ala Leu Asn  
 260 265 270  
 Ser Ser Val Ala Gly Phe Ser Thr Leu Ala Val Asn Val Asn Asp Phe  
 275 280 285  
 Thr Arg Tyr Val Lys His Pro Lys Thr Pro Tyr Val Gln Met Leu Ile  
 290 295 300  
 Leu Pro Leu Val Ala Ala Val Ser Ala Pro Ile Gly Ile Val Ser Gly  
 305 310 315 320  
 Val Ala Ser Lys Ile Met Tyr Gly Thr Ala Met Trp Asp Pro Leu Gln  
 325 330 335  
 Ile Ala Asn Asn Trp Thr Ser Arg Gly Gly Arg Ala Ala Ala Phe Phe  
 340 345 350

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Met Gly Leu Thr Tyr Leu Val Ser Met Ile Ala Gln Asn Ile Ser Asp  
 355 360 365

Asn Thr Val Ala Ala Ala Asn Asp Leu Leu Tyr Phe Phe Pro Arg Tyr  
 370 375 380

Leu Asp Ile Arg Arg Ala Gln Val Ile Val Ile Ile Gly Ala Trp  
 385 390 395 400

Ala Ile Val Pro Trp Lys Ile Leu Gln Asn Gly Thr Ala Phe Leu Ala  
 405 410 415

Phe Leu Gly Ser Leu Ser Ile Phe Leu Gly Pro Ala Ala Gly Ile Phe  
 420 425 430

Val Ala Asp Lys Phe Lys Asn His His Lys Tyr Asp Ile Asp Glu Phe  
 435 440 445

Tyr Asn Pro Ser Gly Ile Tyr Arg Tyr Asn Lys Leu Gly Leu Asn Trp  
 450 455 460

Arg Ala Leu Ile Ala Phe Leu Cys Ala Cys Val Pro Leu Ile Pro Gly  
 465 470 475 480

Met Ala Met Ser Ile Asn Pro Ser Ile Thr Met Pro Asp Gly Val Ile  
 485 490 495

His Leu Tyr Tyr Ile Gly Tyr Phe Tyr Ser Phe Met Thr Ala Phe Leu  
 500 505 510

Ile Tyr Trp Gly Leu Asn Leu Val Phe Pro Ala Lys Glu Thr Leu Leu  
 515 520 525

Glu Glu Ala Val Tyr Pro Pro Lys Ser Asn Ala Glu Leu Val Asp Pro  
 530 535 540

Ser Thr Leu Ser Gly Lys Asp Lys Phe Trp Tyr Tyr Ile Asp Tyr  
 545 550 555

&lt;210&gt; SEQ ID NO 28

&lt;211&gt; LENGTH: 579

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Kluyveromyces lactis

&lt;400&gt; SEQUENCE: 28

Met Ala Gly Val Leu Gly Lys Leu His Asn Leu Leu Val Leu Asp Glu  
 1 5 10 15

Ser Asp Arg Thr Ser Asn Lys Asp Leu Val Pro Met Pro Val Ser Arg  
 20 25 30

Arg Lys Trp Gly Ile Tyr Gly Phe Thr Ser Tyr Trp Thr Leu Leu Cys  
 35 40 45

Leu Cys Ile Ser Thr Trp Ser Gly Gly Ser Ala Leu Leu Leu Tyr Asp  
 50 55 60

Val Gly Thr Asp Gly Glu Leu Thr Leu Ser Gly Met Asn Gly Arg Gln  
 65 70 75 80

Thr Ile Gly Cys Ile Val Leu Ala Asn Phe Phe Ile Ser Ile Ala Ala  
 85 90 95

Ile Ile Asn Ser Val Tyr Gly Ser Glu Tyr His Ile Gly Tyr Ser Val  
 100 105 110

Phe Gln Arg Ile Ile Phe Gly Met Arg Gly Ser Ser Phe Gly Val Leu  
 115 120 125

Ile Arg Ala Ile Leu Ser Val Val Trp Phe Ala Ser Gln Ala Trp Leu  
 130 135 140

Gly Gly Leu Cys Val Asn Val Ile Ile Ser Ser Trp Ser Glu Thr Tyr  
 145 150 155 160

Leu Asn Leu Pro Asn Thr Phe Pro Glu Ser Val Pro Met Thr Arg Gln  
 165 170 175

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Glu Leu Ile Gly Phe Val Ile Phe Leu Val Ile Asn Thr Pro Val Leu  
 180 185 190  
 Met Ile Arg Pro Glu Tyr Phe Asp His Ile Leu Ala Leu Gly Ser Phe  
 195 200 205  
 Cys Met Phe Phe Val Gly Leu Gly Ile Thr Ile Trp Ala Val Thr Ile  
 210 215 220  
 Asn Gly Gly Ser Asn Gly Pro Leu Leu Thr Ala Lys Val Thr Ala Ser  
 225 230 235 240  
 Ser Ser Asp Leu Ala Trp Ser Trp Ile Thr Asn Leu Asn Ala Trp Tyr  
 245 250 255  
 Ser Phe Ile Ile Ala Gly Ile Ser Asn Gln Ser Asp Phe Ser Arg Phe  
 260 265 270  
 Asn Lys Arg Pro Arg Ser Ala Tyr Ile Gly Ile Leu Ile Gly Val Asn  
 275 280 285  
 Val Met Gly Ile Val Leu Pro Leu Met Gly Ile Val Thr Ala Ser Ala  
 290 295 300  
 Leu Leu Glu Lys Tyr Gly Glu Ser Phe Trp Met Pro Asn Asp Ile Cys  
 305 310 315 320  
 Met Tyr Trp Met Gln Leu Asn Tyr Thr Pro Lys Ser Arg Ala Ala Ala  
 325 330 335  
 Phe Phe Ala Gly Leu Gly Leu Leu Ile Ser Gln Leu Gly Val Asn Cys  
 340 345 350  
 Ile Ser Asn Ala Ile Ser Gly Gly Met Asp Leu Ala Ser Ile Phe Pro  
 355 360 365  
 Arg Tyr Ile Asn Ile Arg Arg Gly Ser Ile Leu Ile Met Leu Leu Ala  
 370 375 380  
 Trp Pro Thr Gln Pro Trp Leu Phe Tyr Asn Ala Thr Ser Thr Phe Leu  
 385 390 395 400  
 Thr Val Met Ser Ser Phe Thr Val Phe Ile Thr Pro Leu Thr Ala Met  
 405 410 415  
 Phe Val Cys Asp Tyr Phe Val Ile Arg Lys Gly Val Ile Lys Leu Ser  
 420 425 430  
 Asp Cys Tyr Asp Asp Ser Pro Ser Ser Ile Tyr Trp Phe Gln Tyr Gly  
 435 440 445  
 Ile Asn Trp Lys Asn Ile Leu Cys Phe Leu Cys Gly Ala Ala Pro Gly  
 450 455 460  
 Leu Pro Gly Leu Ile Asn Ala Ala Asn Pro Asn Ile Pro Ile Asn Thr  
 465 470 475 480  
 Gly Ile Glu His Phe Phe Gln Gly Ser Phe Ile Phe Gln Phe Ala Val  
 485 490 495  
 Thr Phe Ala Leu Tyr Tyr Ile Ile Asn Thr Ile Phe Lys Pro Thr Val  
 500 505 510  
 Gly Glu Thr Asp Gln Ile Asp Tyr Tyr His Thr Phe Thr Glu Arg Glu  
 515 520 525  
 Leu Gln Glu Lys Asn Met Ile Ala Asp Asp Gln Glu Asp Ile Gly Val  
 530 535 540  
 His Ser Ile Asp Ser Ser Asn Tyr Leu Ala Asp Pro Thr Glu Val Ser  
 545 550 555 560  
 Leu Arg Asn Leu Lys Ile Asn Ser Pro Glu Lys Ser Asp Ser Thr Asp  
 565 570 575  
 Val Val Leu

&lt;210&gt; SEQ ID NO 29

&lt;211&gt; LENGTH: 543

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&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Aspergillus oryzae

&lt;400&gt; SEQUENCE: 29

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Met Gly Arg Phe Ser Gly Ala Ala Ser Arg Leu His Arg Thr Leu Gln
 1             5             10             15

Leu Ser Gly Asp Thr Asp Gln Trp Arg Asn Arg Asp Leu Ile Pro Leu
      20             25             30

Pro Pro Asp Arg Thr Thr Trp Ser Ser Trp Asp Phe Leu Tyr Leu Trp
      35             40             45

Ser Thr Val Phe Phe Thr Thr Phe Gly Trp Gln Ile Thr Ser Ser Leu
      50             55             60

Leu Gly Leu Gly Leu Asn Val Trp Gln Ser Ile Leu Cys Asn Ile Ile
      65             70             75             80

Thr Lys Phe Leu Gln Thr Ala Val Val Phe Cys Val Ala Trp Pro Gly
      85             90             95

Gly Val Trp His Ile Gly Phe Thr Val Asn Ser Arg Ser Val Phe Gly
      100            105            110

Met Trp Gly Ser Tyr Val Pro Val Ile Leu Arg Ile Phe Leu Cys Ile
      115            120            125

Ile Trp Tyr Gly Val Gln Ala Phe Thr Gly Gly Gln Leu Val Ala Ile
      130            135            140

Ile Leu Ser Thr Ile Phe Ser Gly Tyr His His Met Glu Asn Thr Leu
      145            150            155            160

Pro Glu Ser Ala His Met Thr Thr Lys Gln Phe Val Gly Tyr Val Ile
      165            170            175

Phe Asn Ile Ile Ser Leu Gly Leu Leu Trp Val Pro Pro Asp Lys Leu
      180            185            190

Lys Lys Pro Phe Lys Leu Ile Ala Ala Ile Asn Leu Leu Val Ile Leu
      195            200            205

Gly Leu Ala Ile Gly Leu Ile Ala Gly Ala Arg Gly Gly Ser Leu Gly
      210            215            220

Thr Leu Gln Thr Ser Gln Arg Thr Asp Asn Leu Gly Trp Thr Phe Ile
      225            230            235            240

His Gly Phe Ala Val Val Phe Ser Gly Asn Ala Val Gly Met Ala Ser
      245            250            255

His Ser Asp Phe Ser Arg Phe Ala Arg Arg Pro Gly Ala Gln Val Lys
      260            265            270

Gly Gln Leu Phe Ser Phe Leu Ile Ser Gly Asn Val Val Pro Ile Leu
      275            280            285

Gly Ile Phe Gly Thr Ala Ala Ala Ala Lys Met Tyr Gly Asp Val Asn
      290            295            300

Glu Leu Gly Leu Trp Asn Pro Pro Asn Ile Leu Gln Met Trp Leu Asp
      305            310            315            320

Asn Gln Tyr His Asn Lys Ala Met Arg Ala Ala Ala Phe Phe Val Ala
      325            330            335

Phe Gly Leu Thr Ser Ser Ile Met Ala Met Asn Ser Ile Glu Asn Gly
      340            345            350

Val Ser Gly Gly Met Asp Ile Ala Gly Leu Tyr Pro Arg Tyr Phe Asn
      355            360            365

Ile Arg Arg Gly Ser Tyr Leu Leu Ala Ala Ile Ser Val Val Ile Asn
      370            375            380

Pro Trp Gln Ile Ile Ala Asn Gly Ala Ile Phe Thr Asn Thr Leu Asn
      385            390            395            400

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Ser Phe Gly Val Ile Leu Phe Pro Leu Met Gly Thr Met Val Ala Asp  
 405 410 415

Tyr Tyr Val Val Arg Lys Gln Lys Leu Lys Leu Ser Asp Leu Tyr Arg  
 420 425 430

Ala Asp Ala Ser Ser Ile Tyr Trp Phe Glu Gly Gly Phe Asn Trp Arg  
 435 440 445

Ala Phe Thr Ala Trp Leu Val Gly Phe Ala Pro Ser Val Pro Gly Leu  
 450 455 460

Ala Ala Leu Asn Pro His Asn Thr Gly Ile Pro Ile Gly Leu Thr Tyr  
 465 470 475 480

Thr Phe Tyr Leu Trp Pro Ile Ala Gly Phe Phe Ala Ser Phe Val Leu  
 485 490 495

His Ala Gly Leu Cys Tyr Leu Ser Pro Pro Ala Gly Ile Gly Lys Val  
 500 505 510

Asp Glu Gln Glu Phe His Asp Pro Met Tyr Ser Glu Arg Ser Asp Glu  
 515 520 525

Met Gln Ser Gln Thr Ile Thr Ala Met Glu Lys Gly Gln His Arg  
 530 535 540

<210> SEQ ID NO 30  
 <211> LENGTH: 575  
 <212> TYPE: PRT  
 <213> ORGANISM: Pichia stipitis

<400> SEQUENCE: 30

Met Asp Ala Leu Lys Lys Leu Asp Lys Trp Ile Ala Val Glu Asp Thr  
 1 5 10 15

Ser Thr Glu Arg Gly Glu Asp Gln Ile Arg Ser Asn Glu Asp Leu Asp  
 20 25 30

Pro Thr Pro Ser Asp Arg Arg Thr Trp Lys Met Tyr Asn Tyr Ile Leu  
 35 40 45

Ile Trp Ala Gln Ser Ala Phe Asn Val Asn Glu Trp Asn Thr Gly Ala  
 50 55 60

Ser Leu Met Lys Ala Ser Gly Leu Pro Tyr Gly Gln Thr Ile Gly Ser  
 65 70 75 80

Ala Ile Phe Ser Ile Phe Val Ala Val Ile Phe Thr Ile Ala Asn Ala  
 85 90 95

Arg Ala Gly Ser Thr Tyr His Ile Gly Tyr Pro Thr Leu Ala Arg Ala  
 100 105 110

Thr Phe Gly Val Tyr Gly Ala Tyr Phe Phe Val Ala Ala Arg Gly Phe  
 115 120 125

Val Ala Ile Ile Trp Phe Ser Val Gln Ser Tyr Tyr Gly Ser Met Cys  
 130 135 140

Leu Asp Val Ala Leu Arg Cys Met Phe Gly His Lys Trp Leu Asp Leu  
 145 150 155 160

Lys Asn His Leu Pro Ala Ser Ala Asp Val Gln Ser Arg Ile Leu Leu  
 165 170 175

Ala Phe Phe Leu Phe Trp Leu Ile Gln Phe Pro Leu Met Phe Val His  
 180 185 190

Pro Arg Gln Ile Arg His Phe Phe Thr Val Lys Ser Phe Val Leu Pro  
 195 200 205

Cys Ala Thr Ile Gly Leu Leu Ile Phe Cys Val Lys Lys Gly His Gly  
 210 215 220

Pro Gly Asn Tyr Asp Leu Gly Leu Pro Ile Ser Thr Ser Ser Ser Ala  
 225 230 235 240



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Ile Gly Trp Gly Trp Met Ser Val Met Asn Ser Ile Phe Gly Thr Ile	245	250	255
Ser Pro Met Ile Ile Asn Gln Pro Asp Ile Ala Arg Tyr Ala Lys Lys	260	265	270
Pro Ser Asp Thr Ile Leu Pro Gln Ala Ile Gly Phe Val Leu Ala Lys	275	280	285
Ile Met Ile Met Val Val Gly Met Val Ala Thr Ala Ser Ile Tyr Arg	290	295	300
Ser Tyr Gly Glu Val Tyr Trp Asn Met Trp Asp Leu Met Asn Ala Ile	305	310	315
Leu Asp His Ser Trp Asn Ala Gly Ala Arg Thr Gly Val Phe Phe Val	325	330	335
Ala Val Ser Phe Gly Ile Gly Thr Ala Gly Thr Asn Ile Phe Gly Asn	340	345	350
Ser Ile Pro Phe Ala Cys Asp Ile Thr Gly Leu Leu Pro Lys Tyr Phe	355	360	365
Thr Ile Leu Arg Gly Gln Ile Val Val Ala Ile Leu Ala Trp Ala Ile	370	375	380
Val Pro Trp Lys Phe Leu Thr Asp Ala Ala Lys Phe Leu Thr Phe Leu	385	390	395
Gly Ser Tyr Ser Ile Phe Val Gly Pro Ile Leu Gly Cys Met Leu Ala	405	410	415
Asp Tyr Tyr Phe Val Lys Arg Gly Asn Ile His Val Pro Ser Leu Phe	420	425	430
Thr Lys Lys Ser Ser Gly Val Tyr His Tyr Val Tyr Gly Trp Asn Leu	435	440	445
Trp Ala Cys Phe Ala Trp Ala Gly Ala Ala Ser Ile Cys Ile Pro Gly	450	455	460
Leu Tyr Arg Ala Tyr Tyr Pro Glu Ser Leu Ser Ile Ser Ala Thr Arg	465	470	475
Met Tyr Gln Met Gly Tyr Ile Leu Thr Thr Ile Ser Ser Met Val Phe	485	490	495
Tyr Tyr Cys Leu Ser Leu Ile Phe Lys Pro Gln Ile Tyr Pro Glu Ala	500	505	510
His Arg Asp Thr Pro Lys Thr Trp Glu Tyr Met Arg Thr Thr Asp Gly	515	520	525
Phe Phe Glu Asp Asp Ser Pro Ile Gly Lys Val Gly Tyr Phe Gly Ser	530	535	540
Val Asp Val Phe Thr Gly Glu Lys Val Asp Thr Ser Glu Gly Ser Ser	545	550	555
Val Lys Thr Lys Ser Glu Lys Ile Leu Glu Thr Val Ser Ile Val	565	570	575

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What is claimed is:

1. An isolated *Saccharomyces* strain deficient in the expression of genes involved in nicotinamide riboside import and salvage.

2. The *Saccharomyces* strain of claim 1, wherein said strain does not express Nicotinamide Riboside Kinase 1 (NrK1), Uridine Hydrolase 1 (Urh1), Purine Nucleoside Phosphorylase (Pnp1), and Nicotinamide Riboside Transporter 1 (Nrt1).

3. The *Saccharomyces* strain of claim 1, wherein said strain secretes at least 8 mg/L nicotinamide riboside.

4. The *Saccharomyces* strain of claim 1, wherein said fungus is *Saccharomyces cerevisiae*.

55 5. A method for producing nicotinamide riboside comprising culturing the *Saccharomyces* strain of claim 1 in culture medium and recovering nicotinamide riboside from the medium thereby producing nicotinamide riboside.

60 6. The method of claim 5, wherein the culture medium comprises nicotinic acid or nicotinamide.

7. The method of claim 5, wherein the fungal strain is cultured to an optical density of at least 3.

65 8. The method of claim 5, wherein the nicotinamide riboside is recovered by solubilizing nicotinamide riboside from the medium with methanol and subjecting the nicotinamide riboside to column chromatography.

53

9. A method for producing a nicotinamide riboside-supplemented food product comprising providing a fermentable substrate and fermenting the fermentable substrate in the presence of the *Saccharomyces* strain of claim 1 thereby producing a nicotinamide riboside supplemented food product.

54

10. A nicotinamide riboside supplemented food product fermented in the presence of the *Saccharomyces* strain of claim 1.

11. The product of claim 10, wherein said food product is wine, beer, cider, kvass, root beer, soy sauce or bread.

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